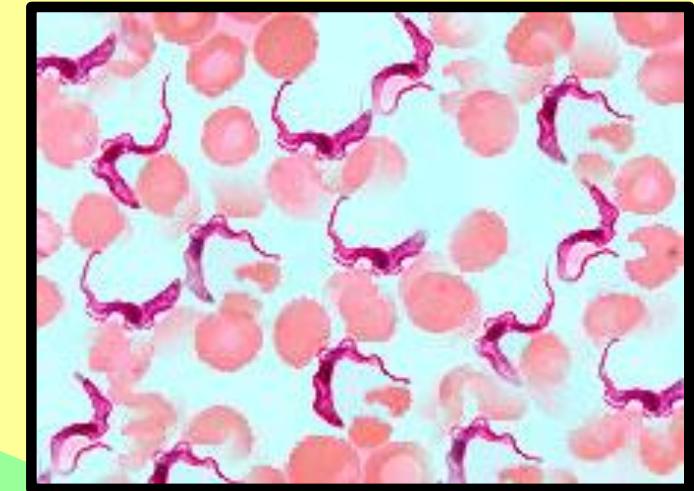
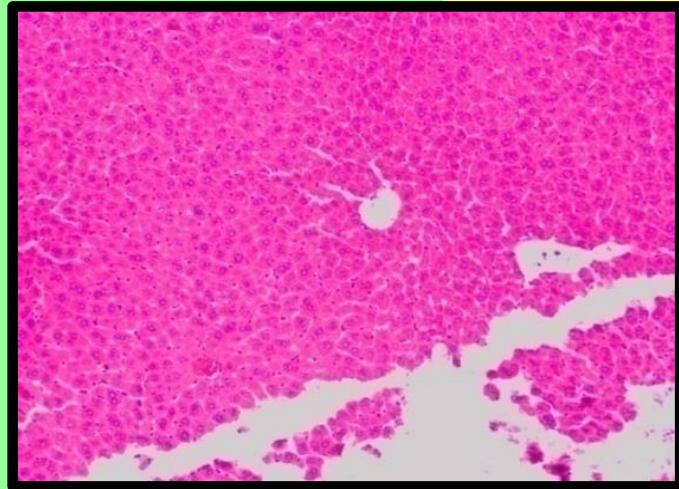


***Piper sarmentosum* Leaf As a Promising Non-toxic Antiparasitic Agent Against *Trypanosoma evansi*-Induced Mice**



Mohd Shukri Baba and Amirul Ikhmal Amir Hamzah

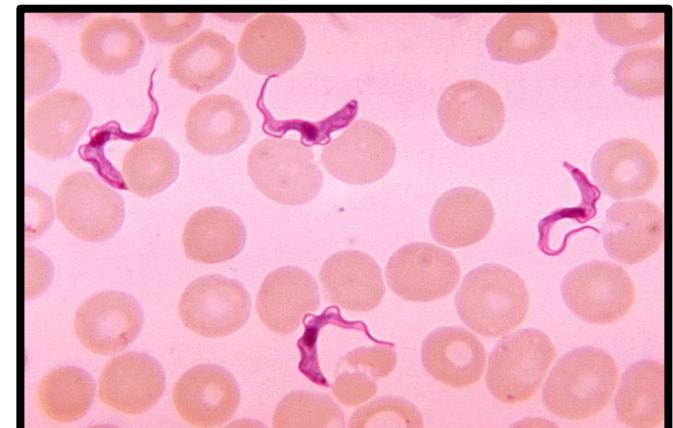
mohd_shukri@iium.edu.my

INTRODUCTION



Trypanosoma evansi

- First discovered by Sir Griffith Evans in Punjab India (1880)
- Haemoflagellated protozoa in both human and animals → zoonotic vector-borne disease
- Caused atypical human trypanosomiasis (AHT) in human and Surra disease in mammals (mostly livestock)
- Wide variety of vectors → worldwide distributed
- Drug resistant in some regions → suramin, pentamidine, berenil



Vectors of *Trypanosoma evansi*



Hirudo medicinalis



Tabanus striatus



Desmodus rotundus



Triatoma infestans



Glossina morsitans



Anthomyia pluvialis

Piper sarmentosum

- A traditional herb & aromatic flowering plant locally known as “kaduk” in Malay
- Wildly & abundant in damp open areas, cleared riverbanks and under shady trees (Seyyedan *et al.*, 2013)
- Well growth on cultivated land in India, Sri Lanka & Southeast Asian region (Hussain *et al.*, 2009).
- Variety of phytochemical constituents & groups identified from various parts of the plants → phenylpropanoids, α -asarone, asaricin, myricetin, sarmentamide A & B, piperitone, naringenin, spathulenol, farnesol, quercetin, etc...



Piper sarmentosum: The Testimonial

Significant antimicrobial activities (Chan & Wong, 2014):

- Antifungal
- Antiamoebic
- Antituberculosis
- Anti-dengue

Antiplasmodial & antileishmanial activities of Pellitorine compound (Souza Oliveira et al, 2018)

The flowers used to treat many chronic diseases (Shim & Gam, 2012):

- Hypertension
- Diabetes mellitus
- Asthma
- Atherosclerosis



Wide range of pharmacological properties (Syed Ab Rahman et al, 2014):

- Wound healing
- Antioxidant
- Anti-inflammatory
- Anti-osteoperosis

Antibacterial activities (Sanusi et al, 2017): *E. coli*, *MRSA*, *B. cereus*, *V. cholera* & *S. typhi*

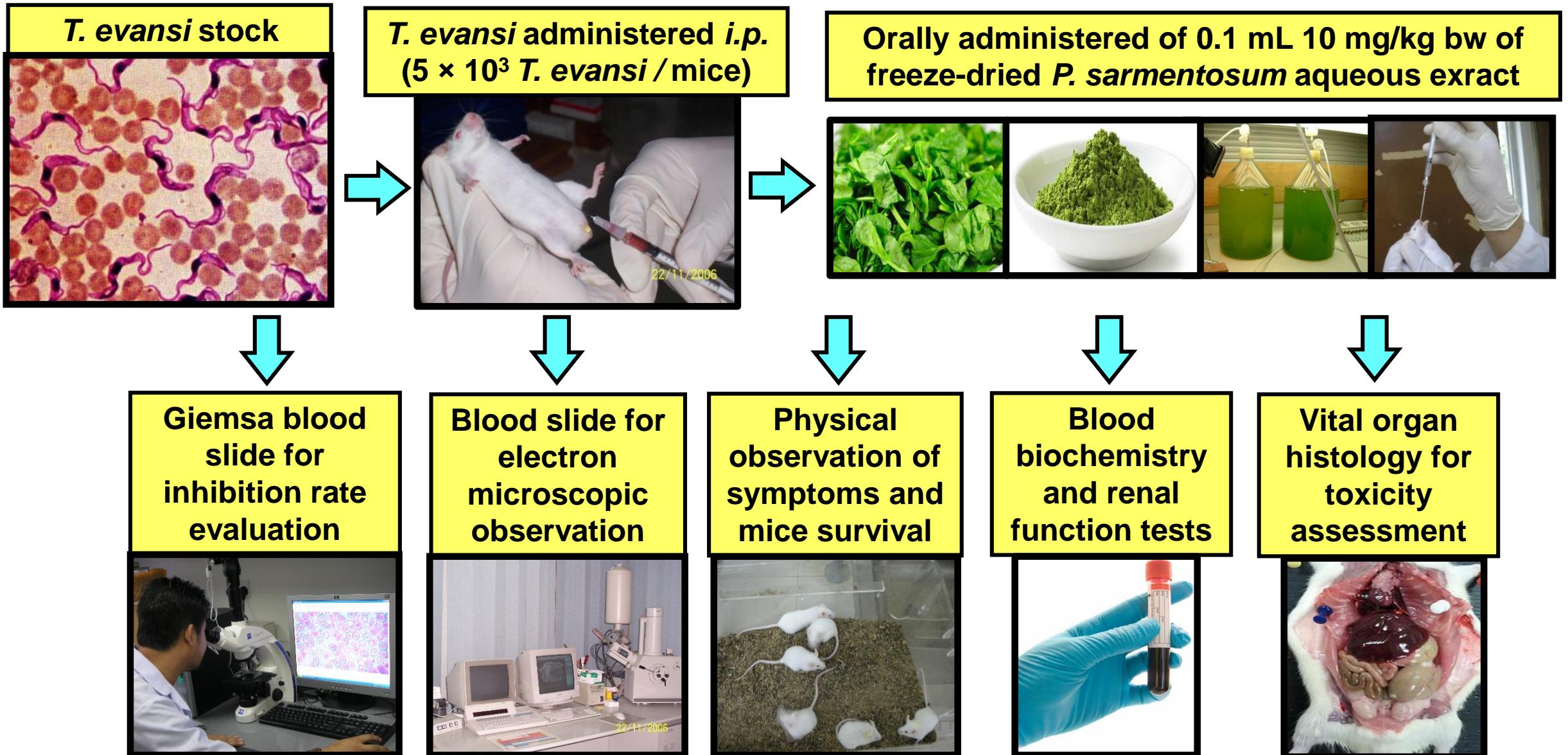
Herbal remedies for many illnesses (Atiax et al, 2011):

- Feet dermatitis
- Toothache
- Headaches
- Coughs

MATERIALS & METHODS



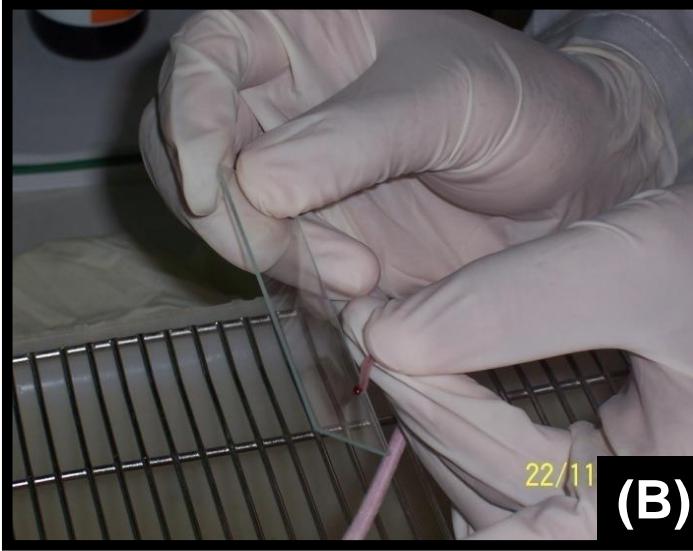
Flow Chart



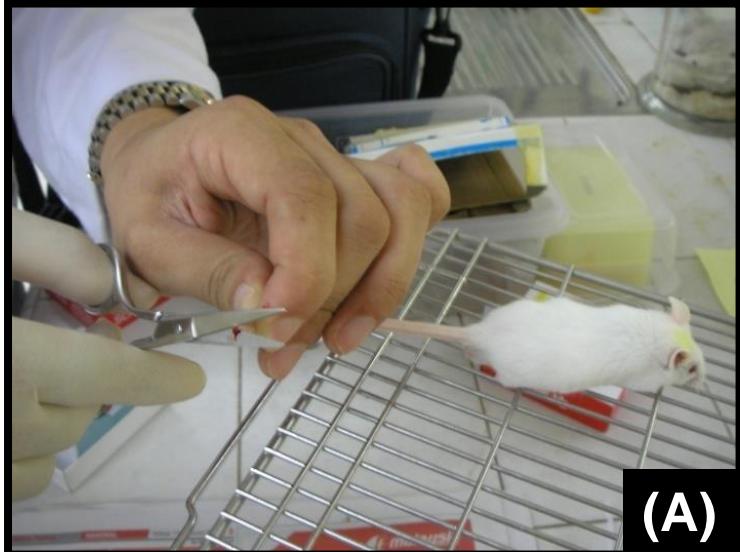
Experimental Design

GROUP	REGIMENS	CODE	DESCRIPTION	<i>P. sarmentosum</i> DOSAGE
TREATMENT	PREVENTIVE	PRE14	14 days pre-infection	0.1 mL 10 mg/kg bw sdH ₂ O-extract
		PRE7	7 days pre-infection	0.1 mL 10 mg/kg bw sdH ₂ O-extract
		PRE3	3 days pre-infection	0.1 mL 10 mg/kg bw sdH ₂ O-extract
	CURATIVE	CUR3	3 days post-infection	0.1 mL 10 mg/kg bw sdH ₂ O-extract
		CUR5	5 days post-infection	0.1 mL 10 mg/kg bw sdH ₂ O-extract
		CUR7	7 days post-infection	0.1 mL 10 mg/kg bw sdH ₂ O-extract
GROUP	REGIMENS	CODE	DESCRIPTION	CONTROL DOSAGE
CONTROL	POSITIVE	POS	Berenil (Sigma-Aldrich KL)	0.01 mL 3.5 mg/kg bw Berenil
	NEGATIVE	NEG	0.9 % Normal Saline	0.1 mL 0.9 normal saline (NS)
	LETHAL	LTN	Infection without treatment	5×10^3 <i>T. evansi</i> / mice (i.p.)

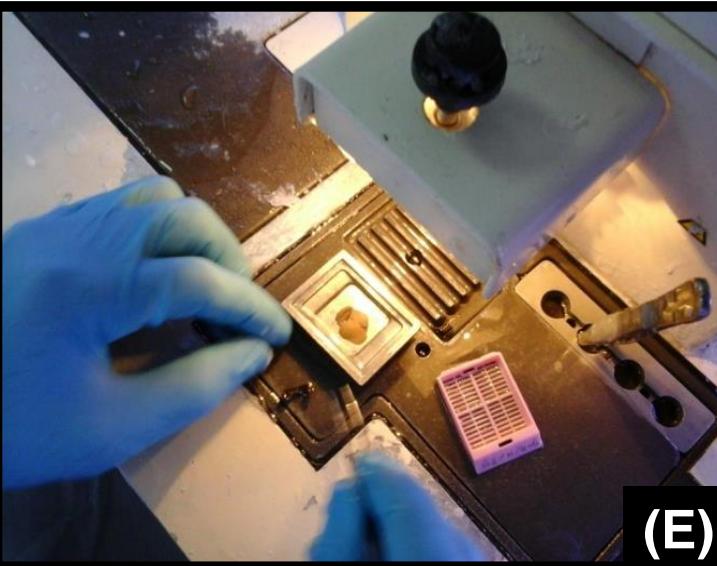
Parasite Administration And Animal Tagging



Giems Staining And Microscopic Observation



Biochemical Test And Histology Of Liver & Kidney

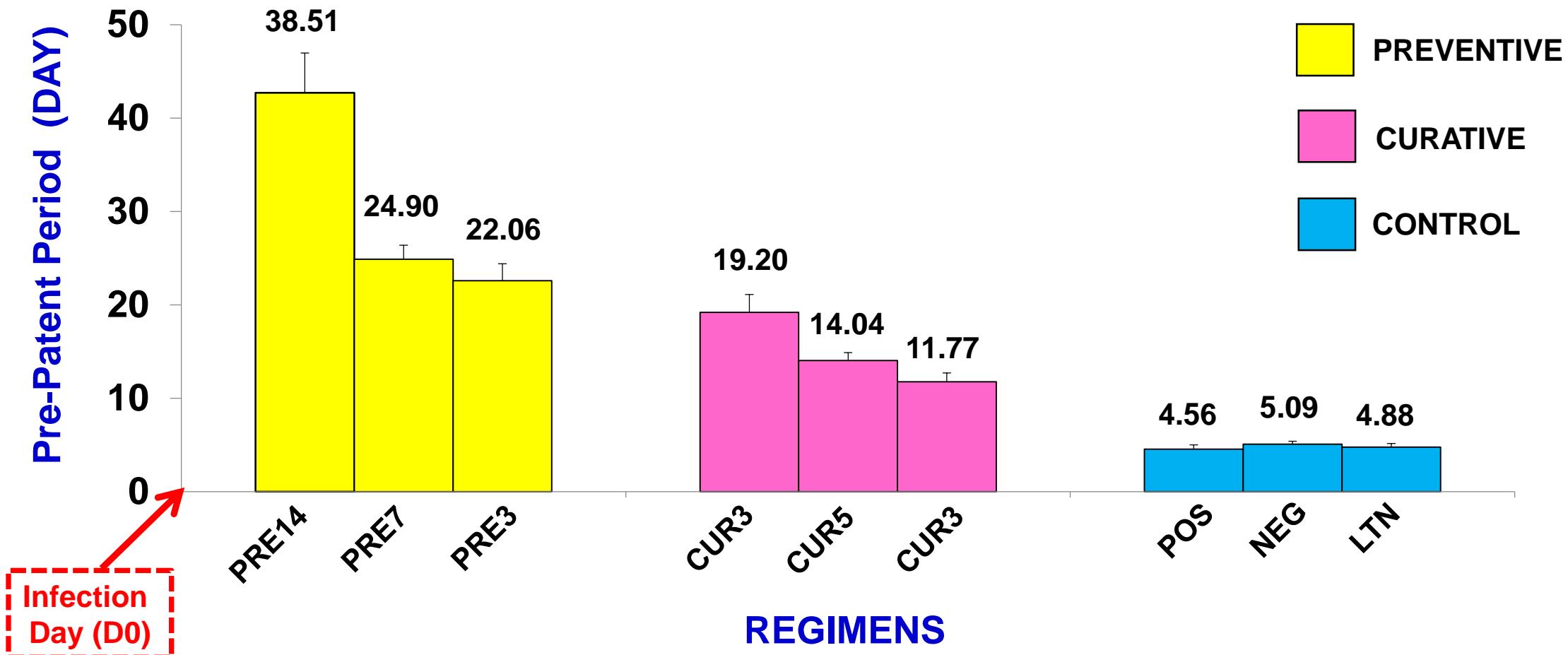


RESULTS & DISCUSSIONS



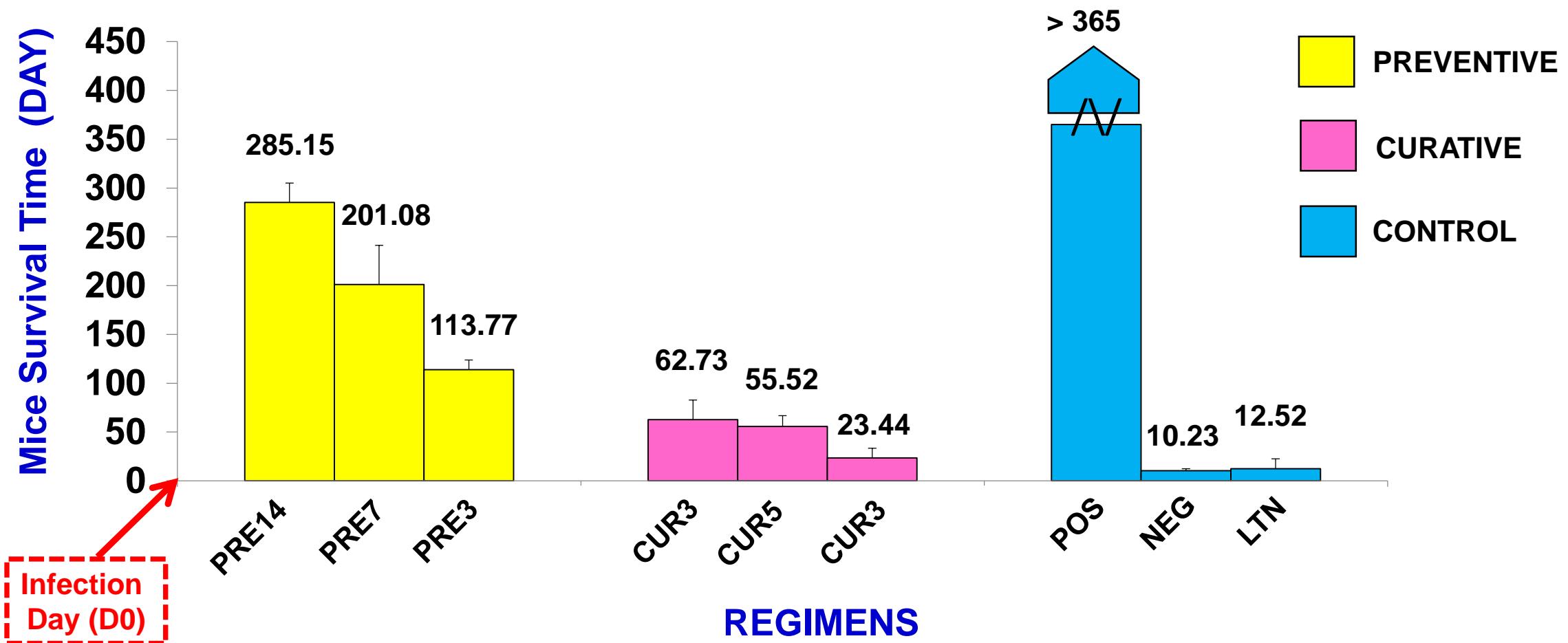
Parasite Pre-Patent Period (Day)

Pre-Patent Period (day) of the mice treated with 0.1 mL 10 mg/kg bw sdH₂O-*P. sarmentosum* extract on D3 post-infection at 5×10^3 *T. evansi*/mice (i.p.) as compared with 3 regimes of control

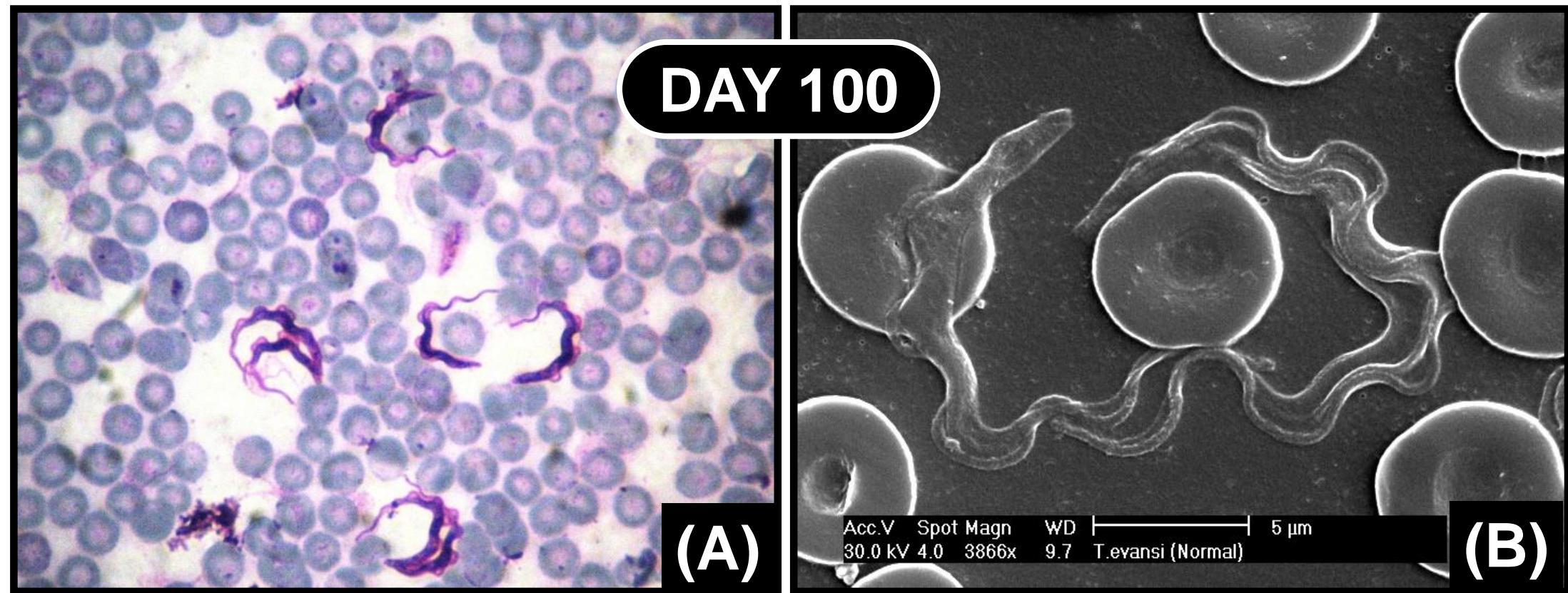


Mice Survival Time (Day)

Mice Survival Time(day) of the mice treated with 0.1 mL 10 mg/kg bw sdH₂O–*P. sarmentosum* extract after infection at 5×10^3 *T. evansi*/mice (i.p.) as compared with 3 regimes of control

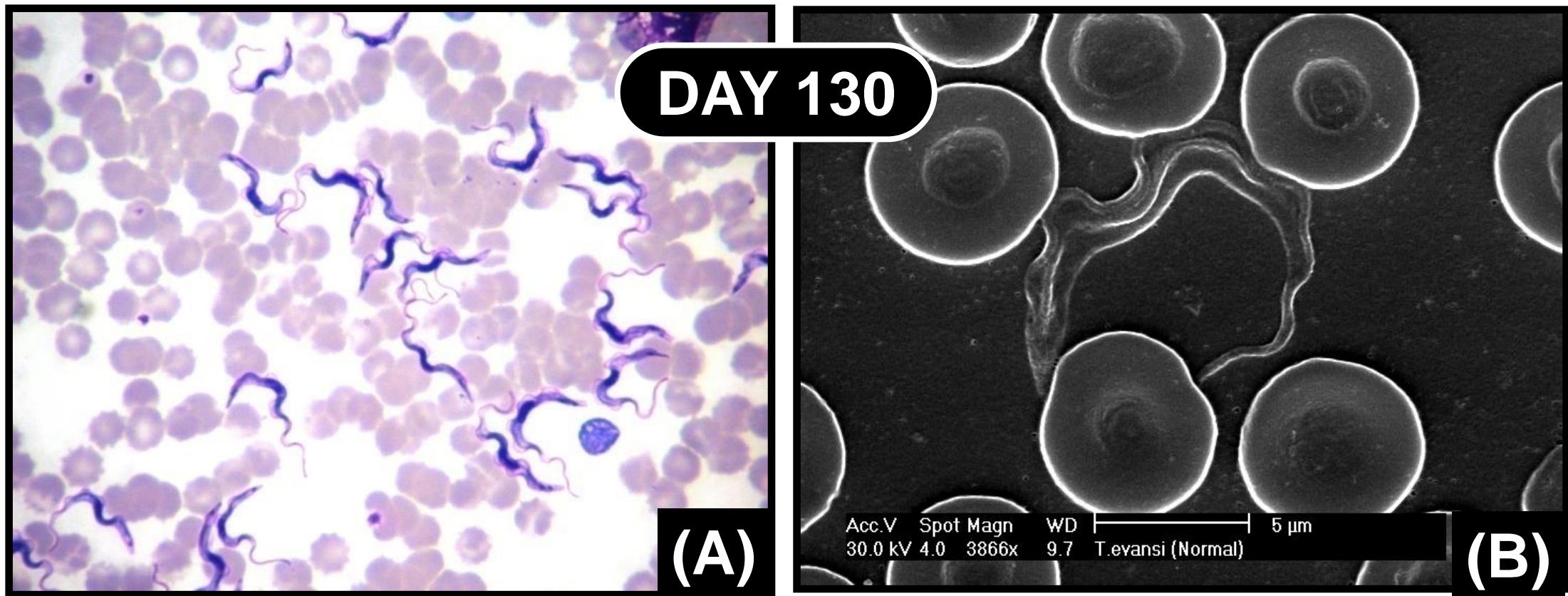


Parasite Survival In PRE14 Mice Group : 100th Day



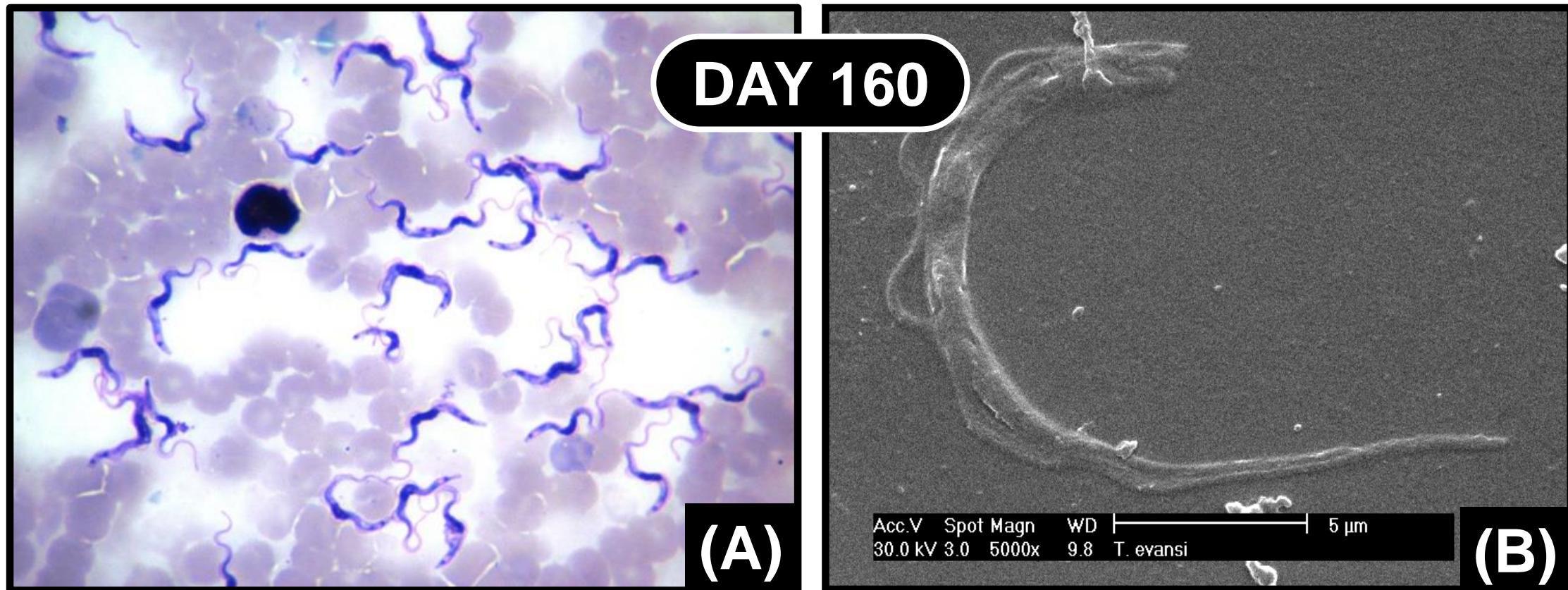
Giemsma thin blood smear of the mice from PRE14 mice group that treated with 0.1 mL 10 mg/kg bw sdH₂O–*P. sarmentosum* extract as observed on day 100 post-infection as observed under x100 magnification of light microscope (A) and x3866 magnification of SEM (Phillips XL30, UK) (B)

Parasite Survival In PRE14 Mice Group : 130th Day



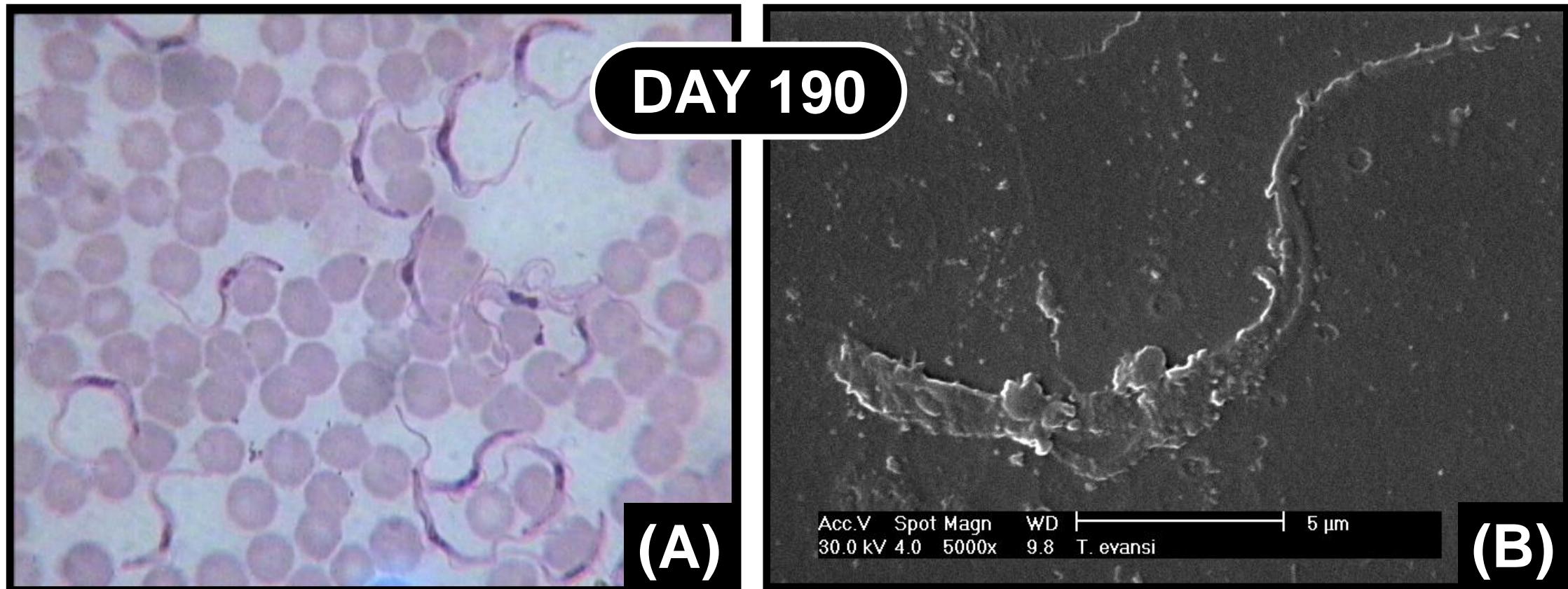
Giemsa thin blood smear of the mice from PRE14 mice group that treated with 0.1 mL 10 mg/kg bw sdH₂O-*P. sarmentosum* extract as observed on day 130 post-infection as observed under x100 magnification of light microscope (A) and x3866 magnification of SEM (Phillips XL30, UK) (B)

Parasite Survival In PRE14 Mice Group : 160th Day



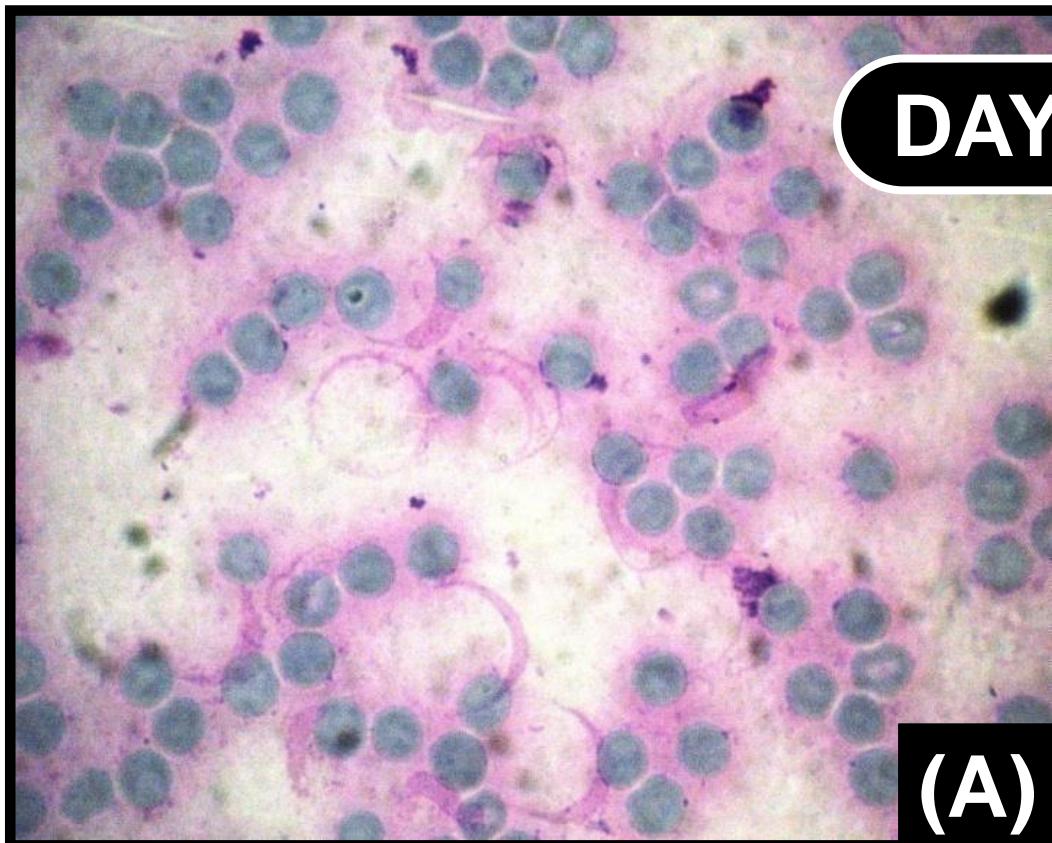
Giemsa thin blood smear of the mice from PRE14 mice group that treated with 0.1 mL 10 mg/kg bw sdH₂O–*P. sarmentosum* extract as observed on day 160 post-infection as observed under x100 magnification of light microscope (A) and x5000 magnification of SEM (Phillips XL30, UK) (B)

Parasite Survival In PRE14 Mice Group : 190th Day



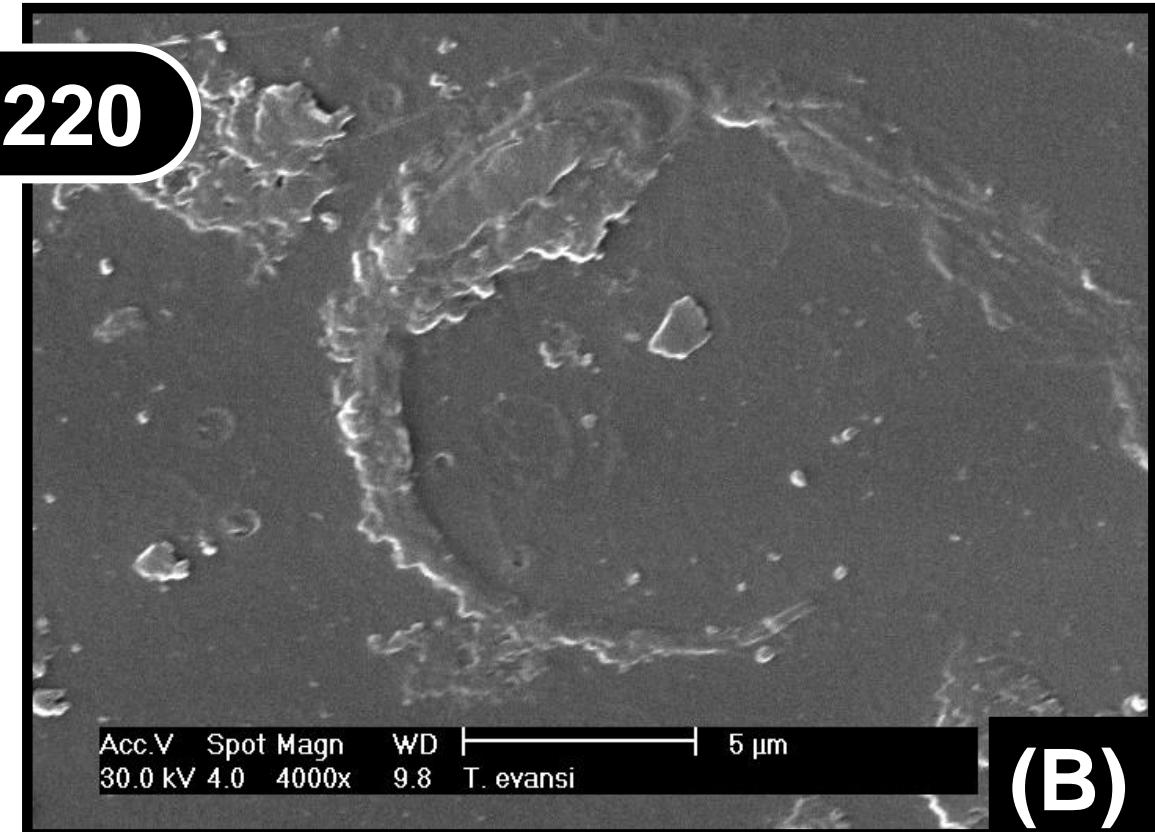
Giemsa thin blood smear of the mice from PRE14 mice group that treated with 0.1 mL 10 mg/kg bw sdH₂O–*P. sarmentosum* extract as observed on day 190 post-infection as observed under x100 magnification of light microscope (A) and x5000 magnification of SEM (Phillips XL30, UK) (B)

Parasite Survival In PRE14 Mice Group : 220th Day



DAY 220

(A)



(B)

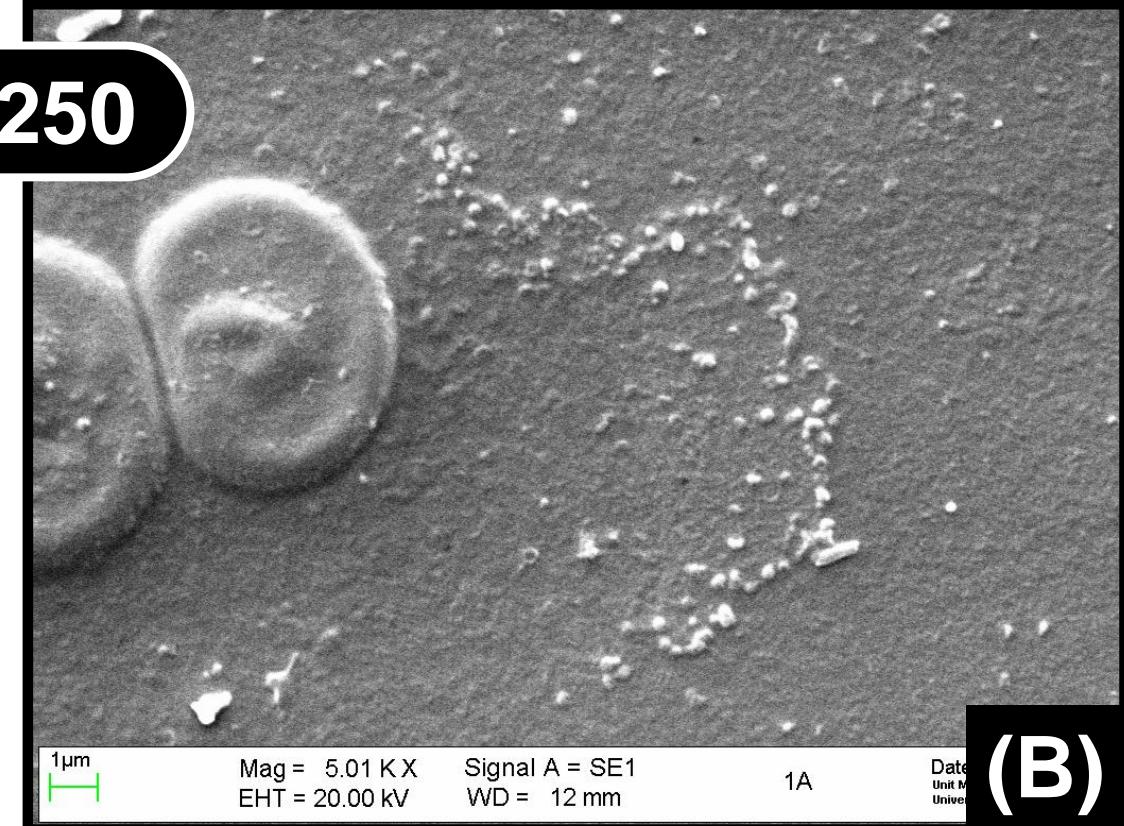
Giemsma thin blood smear of the mice from PRE14 mice group that treated with 0.1 mL 10 mg/kg bw sdH₂O–*P. sarmentosum* extract as observed on day 220 post-infection as observed under x100 magnification of light microscope (A) and x4000 magnification of SEM (Phillips XL30, UK) (B)

Parasite Survival In PRE14 Mice Group : 250th Day



DAY 250

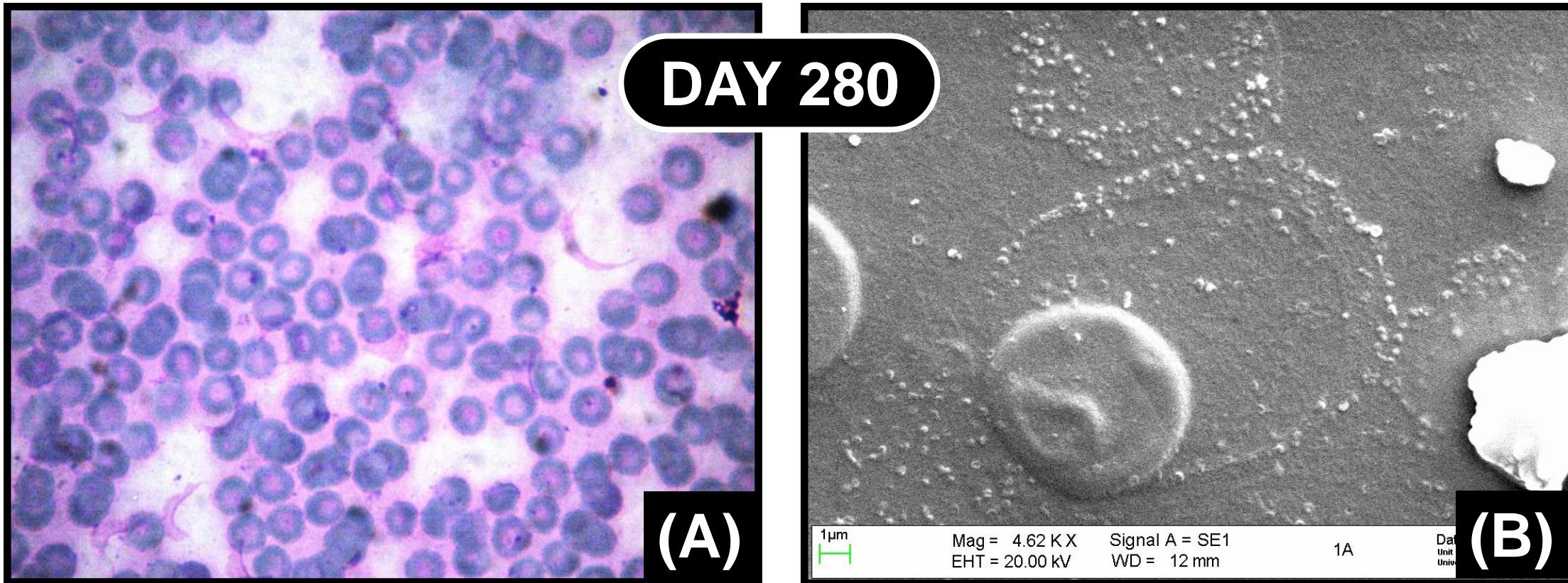
(A)



(B)

Giemsma thin blood smear of the mice from PRE14 mice group that treated with 0.1 mL 10 mg/kg bw sdH₂O–*P. sarmentosum* extract as observed on day 250 post-infection as observed under x100 magnification of light microscope (A) and x5000 magnification of SEM (Leo 1450VP, Japan) (B)

Parasite Survival In PRE14 Mice Group : 280th Day



Giemsma thin blood smear of the mice from PRE14 mice group that treated with 0.1 mL 10 mg/kg bw sdH₂O-*P. sarmentosum* extract as observed on day 280 post-infection as observed under x100 magnification of light microscope (A) and x4600 magnification of SEM (Leo 1450VP, Japan) (B)

Parasite Growth in Berenil-Treated Group (POS)



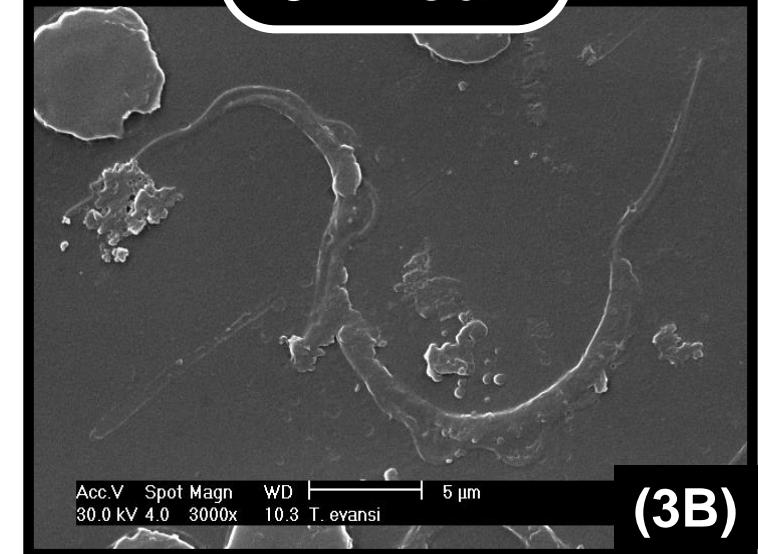
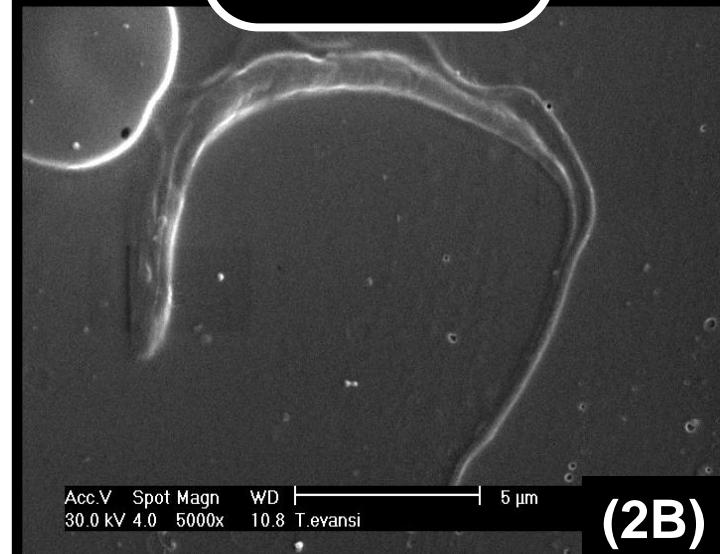
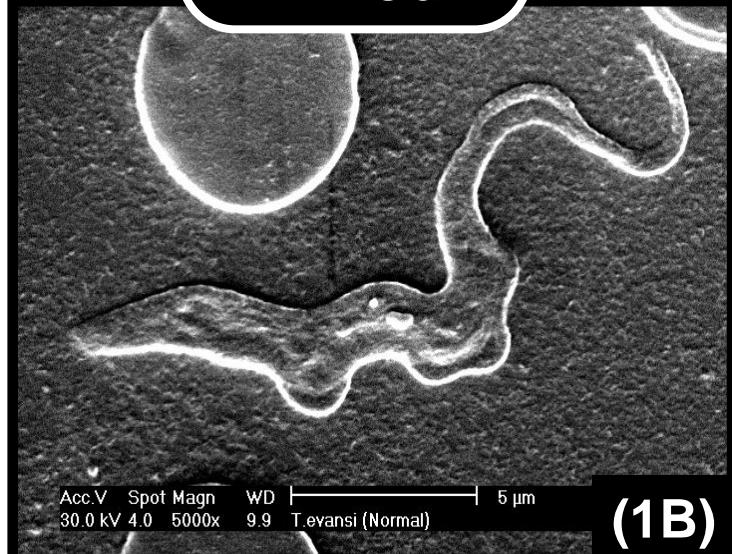
1st hour



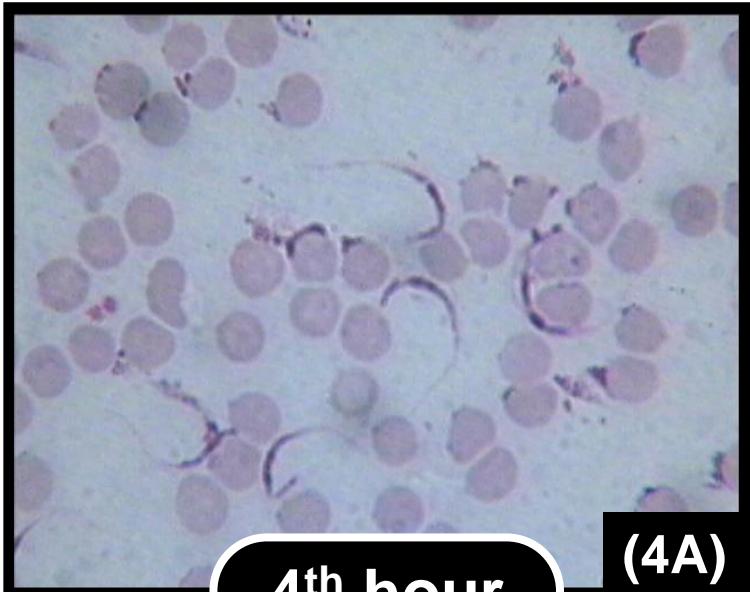
2nd hour



3rd hour



Parasite Growth in Berenil-Treated Group (POS)



4th hour

(4A)



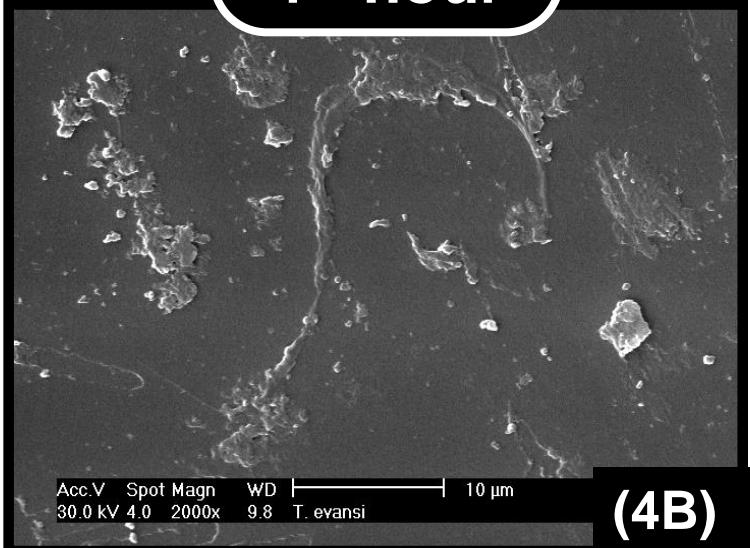
5th hour

(5A)



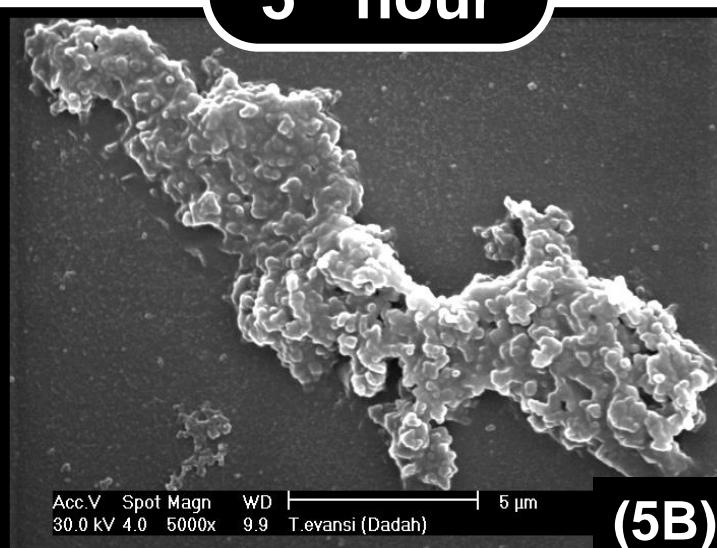
6th hour

(6A)



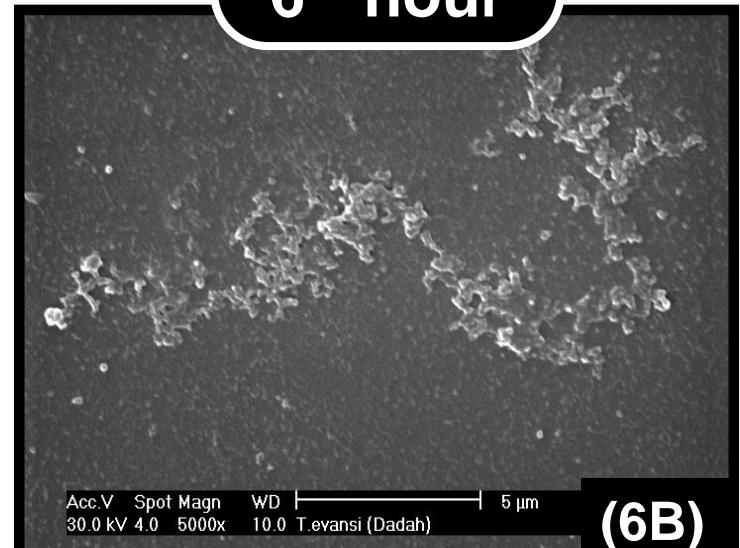
Acc.V 30.0 kV Spot Magn 2000x WD 9.8 T. evansi

(4B)



Acc.V 30.0 kV Spot Magn 5000x WD 9.9 T. evansi (Dadah)

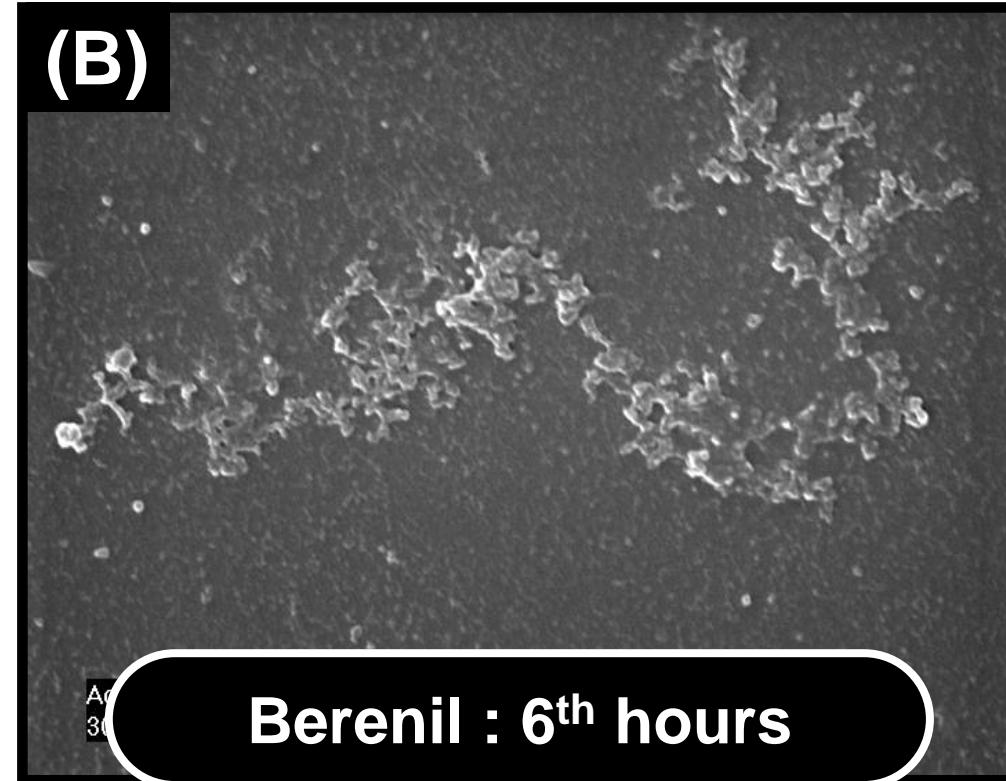
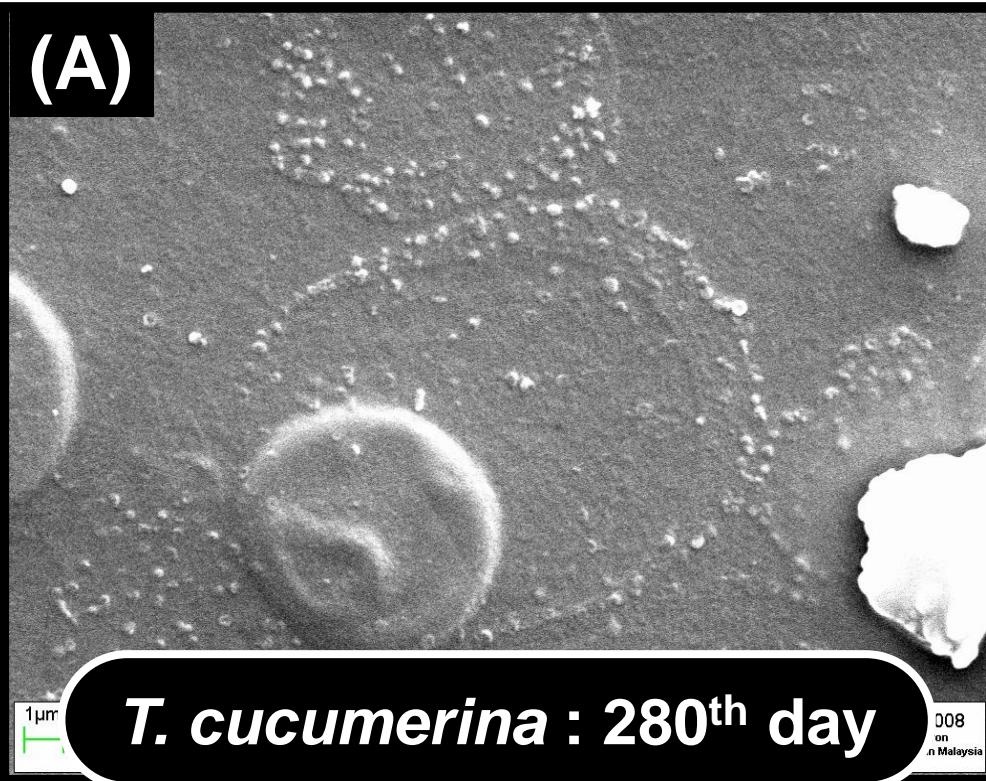
(5B)



Acc.V 30.0 kV Spot Magn 5000x WD 10.0 T. evansi (Dadah)

(6B)

P. Sarmentosum vs Berenil

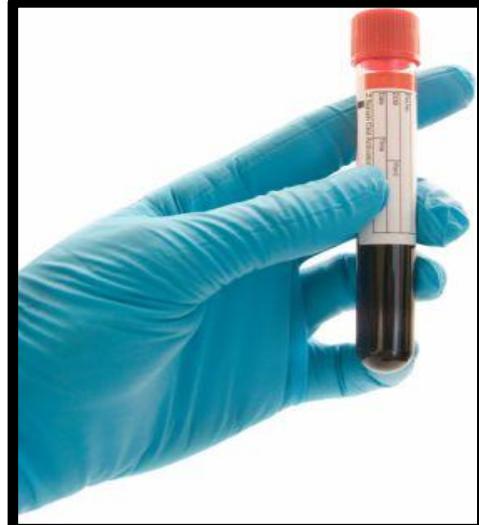


Scanning electron micrograph showed the morphological changes of *T. evansi* in PRE14 mice (0.1 mL 10 mg/kg bw of sdH₂O-*P. sarmentosum* extract) on 280th day post infection (A) and in POS mice at 6th hours post treatment (0.01mL 3.5 mg/kg bw Berenil) (B) as observed under x5000 magnification of SEM

Biochemical Test For Toxicity Assessment



Test	TA	TB	TC	TD	CN	CI	NR	Unit
ALT (*)	41.81 ± 2.14	45.20 ± 1.13	67.57 ± 2.91	90.03 ± 2.02	41.03 ± 3.91	44.83 ± 1.11	40 – 93	IU/L
AST (*)	133.13 ± 2.04	125.93 ± 2.12	167.76 ± 2.27	187.01 ± 2.09	111.62 ± 1.19	134.43 ± 4.01	92 – 206	IU/L
ALP (*)	62.76 ± 2.33	59.4 ± 2.97	69.2 ± 2.90	68.03 ± 2.10	61.46 ± 2.46	58.32 ± 2.97	54 – 115	IU/L
STP (*)	6.12 ± 2.32	7.21 ± 3.81	7.93 ± 2.01	8.83 ± 3.90	6.40 ± 1.01	6.80 ± 3.06	5.8 – 9.5	g/dL



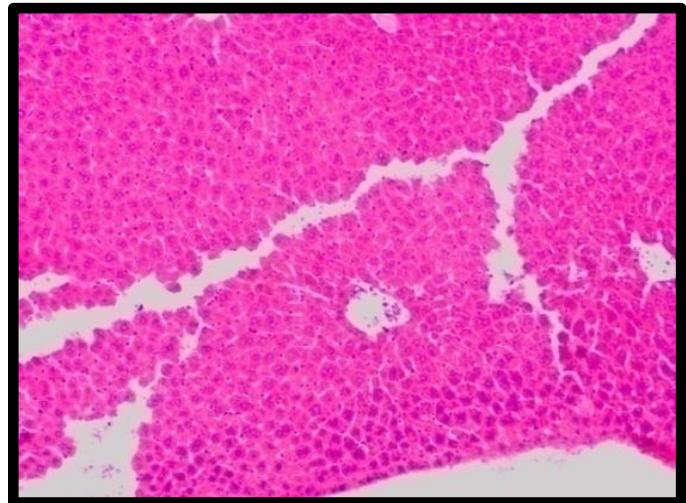
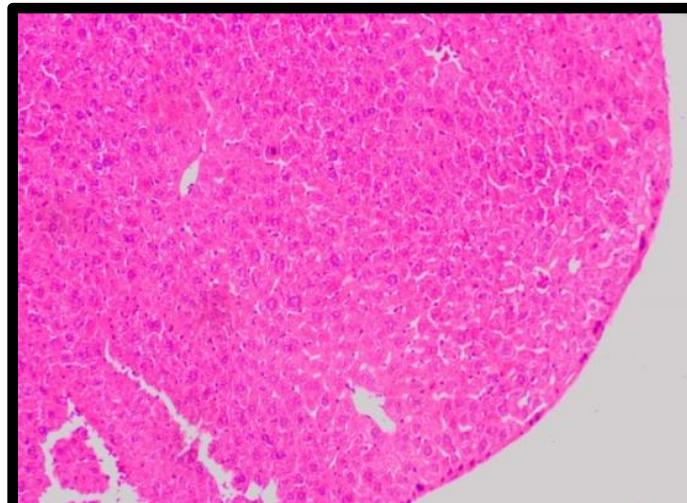
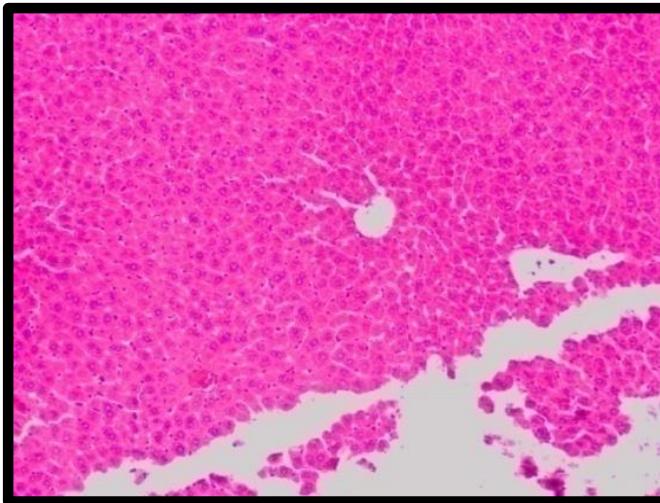
- TA : Sub-acute regime – Daily treatment (28 days)
 TB : Sub-acute regime – Daily treatment (28 days) 2 hours post-infection
 TC : Sub-chronic regime – Daily treatment (90 days)
 TD : Sub-chronic regime – Daily treatment (90 days) 2 hours post-infection
 CN : Control regime – Normal mice without infection and treatment
 CI : Control regime – Infected mice on D0
 ALT : Alanine aminotransferase
 AST : Aspartate transaminase
 ALP : Alkaline phosphatase
 STP : Serum total protein

(*) All values were expressed as mean ± standard errors (se)

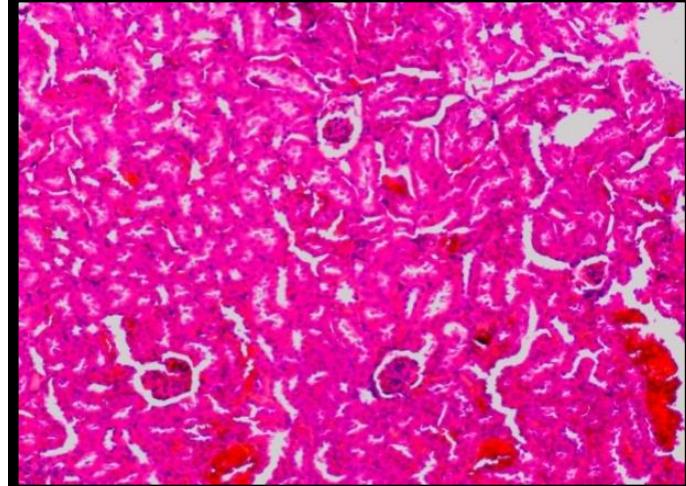
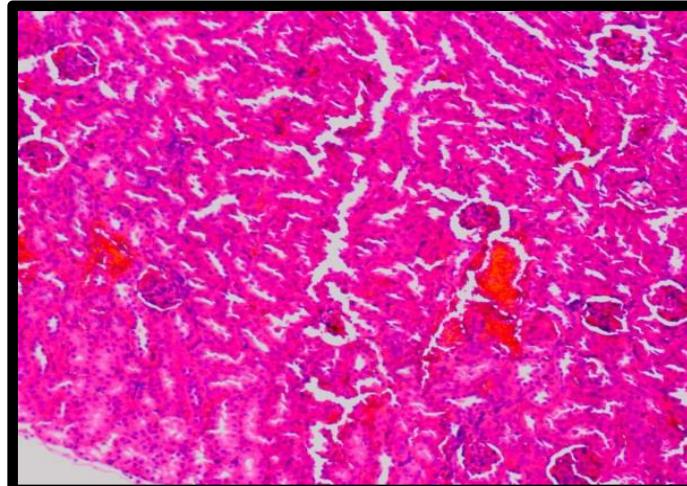
(*) All NR values were referred from Research Animal Resources, University of Minnesota, USA

Organ Histology For Toxicity Assessment

Liver



Kidney



Treatment (Acute)

Treatment (Sub-acute)

Control

CONCLUSIONS



Hypothesis

- Tryptophol toxin and stochastic genetic modification of VSA is still the best ‘weapon’ for *T. evansi* survival (Otto *et al.* 2010).
- New wave of infection & periodic changes of antigenic variation → changes in parasitemia peaks → longer survival time of the parasite & chronic infection on host (Salleh *et al.* 2009)
- The action of pellitorine ($C_{13}H_{25}ON$) molecule in *P. sarmentosum* against –thiol group of parasite enzymes in which crucial for parasite proliferation (Souza Oliveira *et al.*, 2018).
- Bioactive compound of Sarmentamide A in *P. sarmentosum* inhibited the important enzymes (alcohol dehydrogenase, cysteine proteinase and thioredoxin reductase) for the stability of the redox reaction in fungal cells such as *A. fumigatus* & *C. albicans* (Tuntiwachwuttikul *et al.*, 2006)

Future Plans

Various solvents
of *T. cucumerina*
extract

Mechanism
of action

In-vitro
anti-trypanosomal
screening

Concentration- &
time-dependant
alteration

Clinical &
molecular
approaches

Screening
against *T. cruzi*
and *T. brucei*



Absolute Hypothesis

EAT KADUK..!

NO HARM TO EAT AS MUCH AS YOU CAN



Absolute Hypothesis



REFERRENCEES



REFERENCES

- Atiax, E., Ahmad, F., Sirat, H.M and Arbain, D. 2011. Antibacterial Activity and Cytotoxicity Screening of Sumatran Kaduk (*Piper sarmentosum* Roxb.). *Iranian Journal of Pharmacology & Therapeutics* 10: 1-5
- Flávio Augusto de Souza Oliveira, Guilherme Matos Passarini, Daniel Sol Sol de Medeiros, Ana Paula de Azevedo Santos, Saara Neri Fialho, Aurileya de Jesus Gouveia, Marcinete Latorre, Elci Marlei Freitag, Patrícia Soares de Maria de Medeiros, Carolina Bioni Garcia Teles, and Valdir Alves Facundo. (2018). Antiplasmodial and antileishmanial activities of compounds from *Piper tuberculatum* Jacq fruits. *Rev Soc Bras Med Trop* 51(3):382-386
- Shim, S and Gam, L. 2012. Analysis of *Piper sarmentosum* proteome using two dimensional gel electrophoresis and mass spectrometry. *J. Mol. Biol. Biotechnol.* 20 (4) : 124-139.
- Syed Ab Rahman, S. F., Sijam, K. and Omar, D. 2014. Identification and Antibacterial Activity of Phenolic Compounds in Crude Extracts of *Piper sarmentosum* (*Kadok*). *Journal of Pure & Applied Microbiology*, 8 (Supl. Edn. 2) : 483-490.
- Chan, E. W. C and Wong, S.K. 2014. Phytochemistry and Pharmacology of Three *Piper* Species: An Update. *International Journal of Pharmacognosy* 1 (9): 534-544

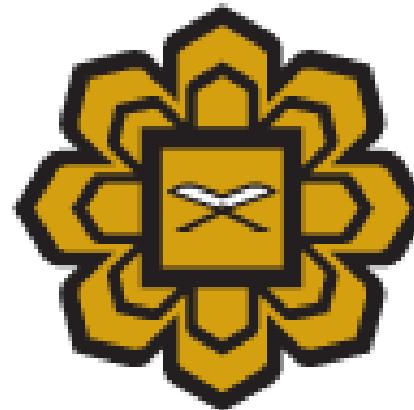
REFERENCES

- Nurul 'Adani Sanusi, Rabiatul Adawiyah Umar, Mohd Nizam Zahary, Mohd Adzim Khalili Rohin,Mohd Ridzuan Pauzi and Salwani Ismail .(2017). Chemical Compositions and Antimicrobial Properties of *Piper Sarmentosum* – A Review. Journal of Dental and Medical Sciences. 16(8): 62-65
- Nok, A.J., William, S. & Onyenekwe, P.C. 1996. *Allium sativum* : induced death of African trypanosomes. *Parasitol. Res.* 82: 634-637
- Denise, C.A., Fabio, D.A., Alejandro, M.K. & Silvia, R.U. 2004. Antileishmanial activity of the terene Nerolidol. Departamento de Parasitologia, Instituto de Ciencias Biomedicas, Universidade de Sao Paulo, Brazil
- Zainal-Abidin, B.A.H. 1992. Infections of *Trypanosoma evansi* in Malaysia. *Malays. Applied Biology* 10: 1-8
- Lazuardi M, 1998. The trypanocide effect of suramin against to *Trypanosoma evansi*. The Indonesian Journal of Parasitology. 11 (1) : 26-32.
- Kaminsky R and Zweygarth A, 1989. Effect of in vitro cultivation on the stability of resistance of *trypanosoma brucei brucei* to diminazene, isometamidium, quinapyramine, and Mel B. *J. Parasitol.*, : 42-45

REFERENCES

- Croft SLJA Urbina, Brun R, 1997. Chemotherapy of human leishmaniasis and trypanosomiasis. In G Hide, Mottram JC, Coombs GH and Holmes PH (eds), Trypanosomiasis and Leishmaniasis. CAB International, Tucson, Ariz. P. 245-247
- Hussain, K., Ismail, Z., Sadikun, A. and Ibrahim, P. 2009. Cytotoxicity Evaluation and Characterization of Chloroform Extract of Leaf of *Piper sarmentosum* Possessing Antiangiogenic Activity. *Pharmacologyonline* 2: 379-391.
- Tuntiwachwuttikul P, Phansa P, Pootaeng-On Y, Taylor WC. (2006). Chemical constituents of the roots of *Piper sarmentosum*. *Chem Pharm Bull*, 54, 149–151.
- Otto, M.A., Da Silva, A.S., Gressler, L.T., Farret, M.H., Tavares, K.C.S., Zanette, R.A., Miletti, L.C., Monteiro, S.G., 2010. Susceptibility of *Trypanosoma evansi* to human blood and plasma in infected mice. *Veterinary Parasitology* 168, 1–4.
- Salleh, M.A., Al-Salhy, B.M., Sanousi, S.A., 2009. Oxidative stress in blood of camels naturally infected with *Trypanosoma evansi*. *Veterinary Parasitology* 162, 192– 199
- Menezes, V.T., Queiroz, A.O., Gomes, M.A., Marques, M.A., Jansen, A.M., 2004. *Trypanosoma evansi* in unbred and Swiss-Webster mice: distinct aspects of pathogenesis. *Parasitology Research* 94, 193–20

ACKNOWLEDGEMENT



الجامعة الإسلامية العالمية ماليزيا
INTERNATIONAL ISLAMIC UNIVERSITY MALAYSIA
يونیورسiti إسلامي انتارا بعثسا ملسيستيا
Garden of Knowledge and Virtue



UPM
UNIVERSITI PUTRA MALAYSIA
BERILMU BERBAKTI



KEMENTERIAN
PENDIDIKAN
MALAYSIA

Thank You



Rationale Of The Study

Reliability of Anti-Trypanosomal Drugs

- Resistant issues in India, Thailand & Indonesia
- Unaffordable → expensive in certain regions
- Wrong dosage & concentration → side effects



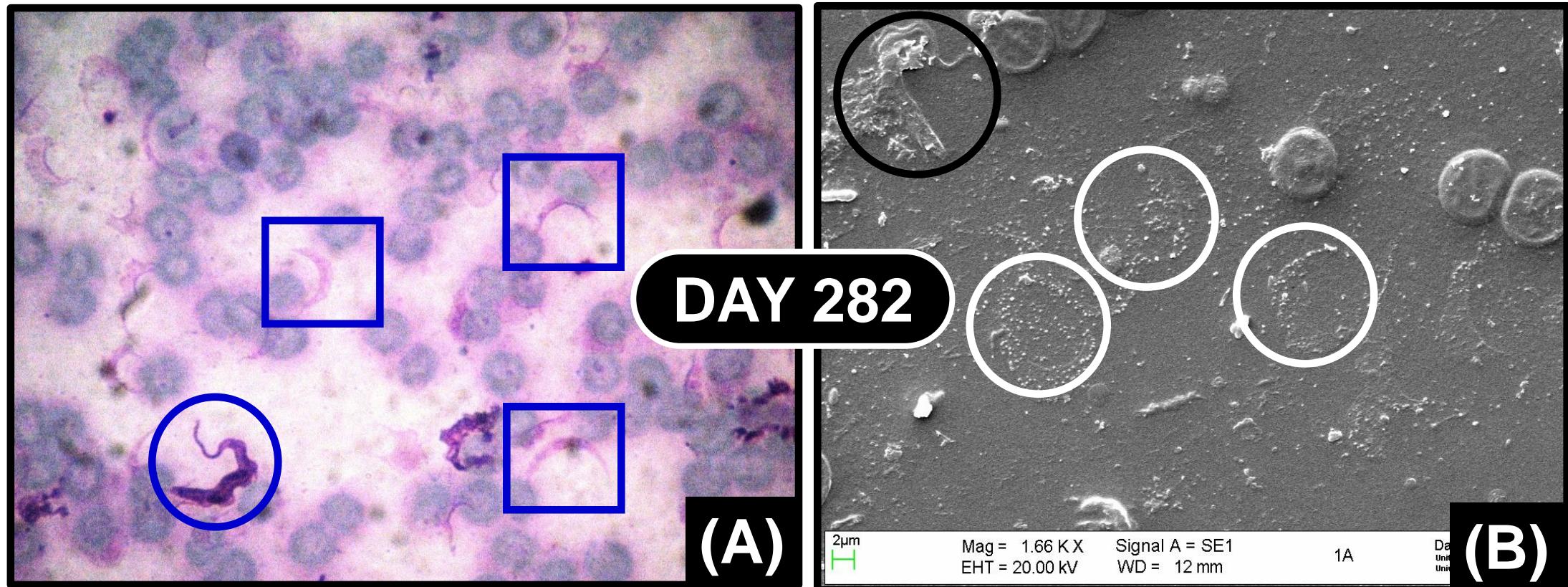
Economic Growth & Biotechnology Sector

- Biotechnology → main focus in the next decade
- Snake gourd → consumable & easily manipulated
- AHT & Surra → influenced productivity of human & livestock

Current Issues of *T. evansi*

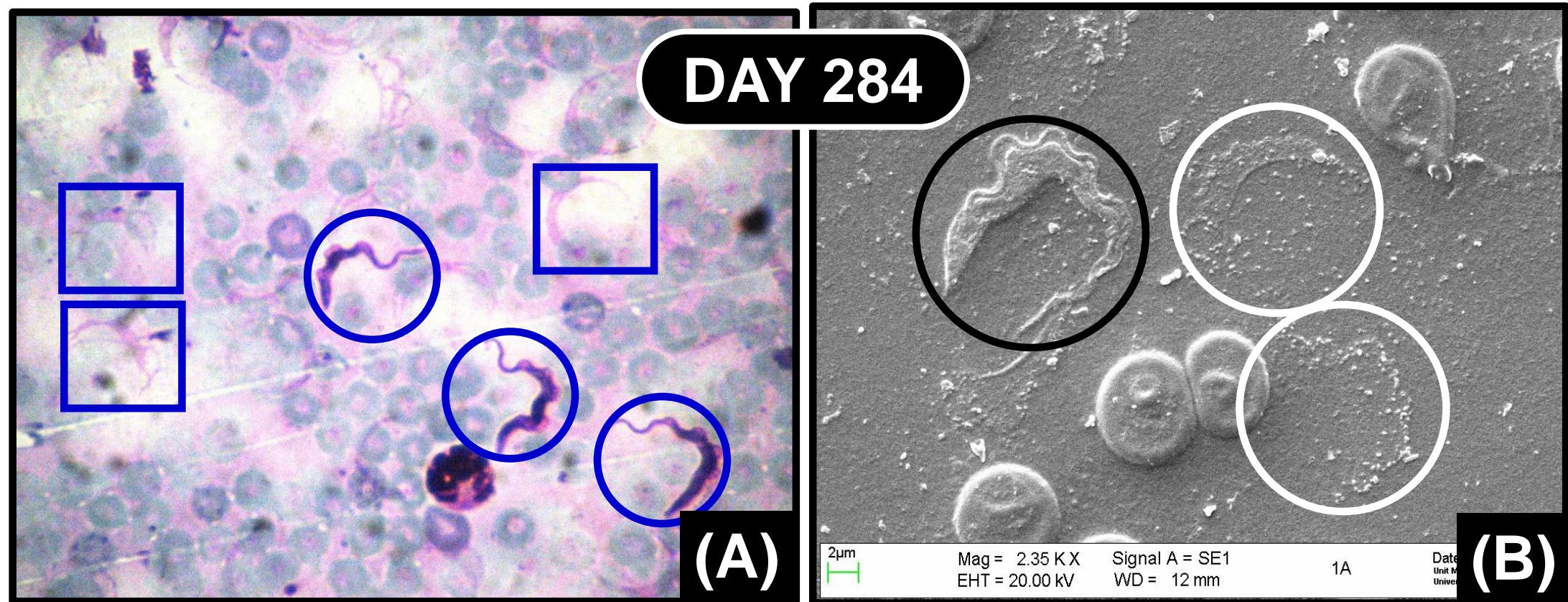
- Trans-host boundary : animal → human (Assam India 2008)

Parasite Survival In PRE14 Mice Group : 282nd Day



Reemerged of *T. evansi* which survived in PRE14 group mice on day 282 due to the action of 'variable surface glycoprotein (VSA) stochastic genetic modification' as observed under x100 magnification of light microscope (A) and x1600 magnification of SEM (Leo 1450VP, Japan) (B).

Parasite Survival In PRE14 Mice Group : 284th Day



Reemerged of *T. evansi* which survived in PRE14 group mice on day 284 due to the action of 'variable surface glycoprotein (VSA) stochastic genetic modification' as observed under x100 magnification of light microscope (A) and x2300 magnification of SEM (Leo 1450VP, Japan) (B). Later the mice died on day 286

Variable Surface Glycoprotein (VSG)

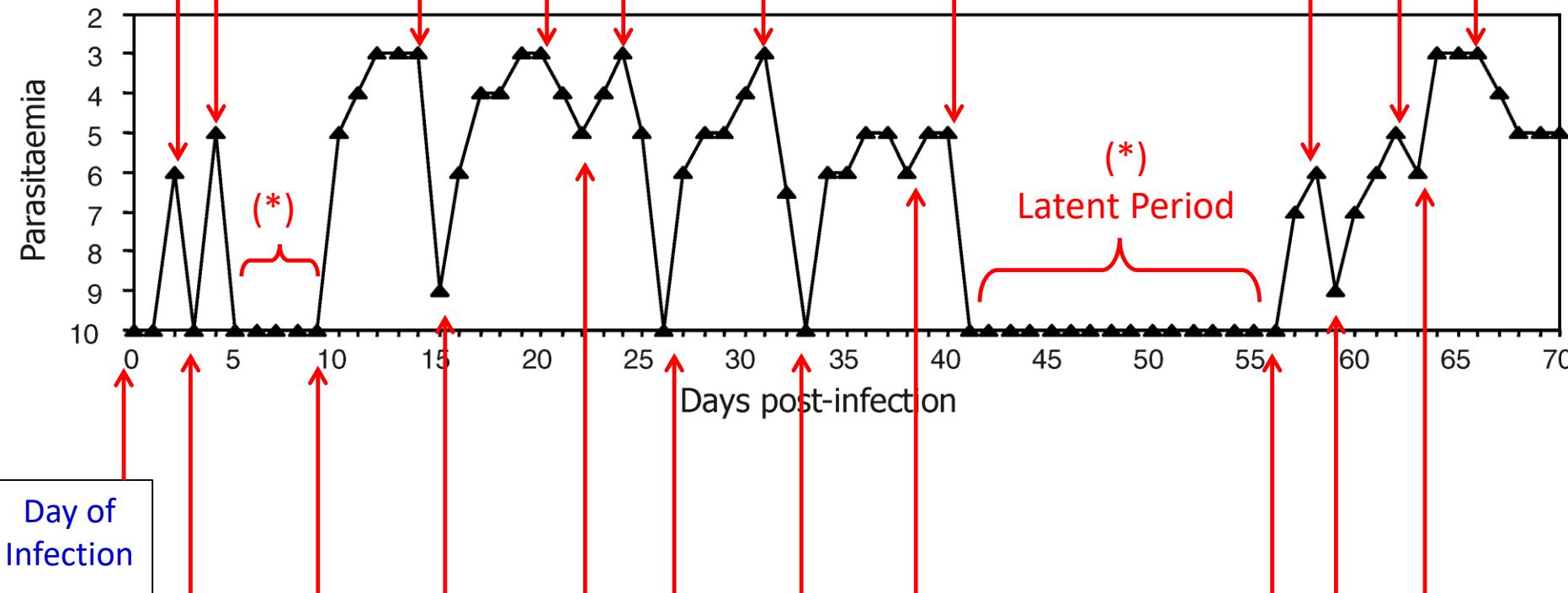
- Survival factor of *Trypanosoma* spp. in the infected host
- High density layer on the parasite cell membrane
- Contained 1×10^9 similar & uniformed glycoprotein molecules expressed by VSG-Trypanosome gene
- Protect the parasite from being identified/action of the host immune system
- Similar & uniformed glycoprotein molecule → only end region of 'N-terminal loops' structure (300-500 amino acid structures) can be identified by the host immune systems → specific antibody-antigen mechanisms

Variable Surface Glycoprotein (VSG)

- When the end region of 'N-terminal loops' structure being identified by the host immune systems → VSG-stochastic genetic modification' of the parasite plays the role.
- VSG stochastic genetic modification = periodic changes of antigenic variation → the structures & characteristics of parasite cell membrane was modified whenever confronted with the host's specific immune system which may varies.
- Periodic changes of antigenic variation → changes in parasitemia waves → longer survival time of the parasite → chronic infection on host

Survival Pattern of the Trypanosomiasis Infected-Host Due to VSG-Stochastic Genetic Modification Phenomenon

Effectuation of the changes in host's specific immune system



Mechanism of Trypanosome VSG-stochastic genetic modification