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A review on chitin dissolution as preparation for electrospinning application

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Abstract

Electrospinning has been acknowledged as an efficient technique for the fabrication of continuous nanofibers from polymeric based materials such as polyvinyl alcohol (PVA), cellulose acetate (CA), chitin nanocrystals and others. These nanofibers exhibit chemical and mechanical stability, high porosity, functionality, high surface area and one-dimensional orientation which make it extremely beneficial in industrial application. In recent years, research on chitin - a biopolymer derived from crustacean and fungal cell wall - had gained interest due to its unique structural arrangement, excellent physical and chemical properties, in which make it biodegradable, non-toxic and biocompatible. Chitin has been widely utilized in various applications such as wound dressings, drug delivery, tissue engineering, membranes, food packaging and others. However, chitin is insoluble in most solvents due to its highly crystalline structure. An appropriate solvent system is required for dissolving chitin to maximize its application and produce a fine and smooth electrospun nanofiber. This review focuses on the preparation of chitin polymer solution through dissolution process using different types of solvent system for electrospinning process. The effect of processing parameters also discussed by highlighting some representative examples. Finally, the perspectives are presented regarding the current application of electrospun chitin nanofibers in selected fields. © 2024 Elsevier B.V.

Author Keywords

Chitin dissolution; Electrospinning; Nanofibers

Index Keywords

Biocompatibility, Biopolymers, Chemical stability, Chitin, Dissolution, Drug delivery, Electrospinning, Solvents, Tissue engineering; Cellulose acetates, Chitin dissolution, Fungal cell walls, High porosity, High surface area, Non-toxic, One-dimensional, Physical and chemical properties, Solvent system, Structural arrangement; Nanofibers; biopolymer, cellulose acetate, chitin, deep eutectic solvent, inorganic acid, inorganic salt, ionic liquid, nanocrystal, nanofiber, organic solvent, polyvinyl alcohol, polymer, solvent; aqueous solution, biocompatibility, biodegradability, crystal structure, dissolution, drug delivery system, electrospinning, food packaging, fungal cell wall, human, nonhuman, Review, scanning electron microscopy, tissue engineering, waste water management, chemistry, procedures, tissue engineering; Application, Chemical Properties, Chitin, Dissolving, Processing, Review, Solvents; Chitin, Polymers, Polyvinyl Alcohol, Solvents, Tissue Engineering

Chemicals/CAS

cellulose acetate, 9004-35-7; chitin, 1398-61-4; polyvinyl alcohol, 37380-95-3, 9002-89-5; Chitin; Polymers; Polyvinyl Alcohol; Solvents

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References

- Refate, A., Mohamed, Y., Mohamed, M., Sobhy, M., Samhy, K., Khaled, O., Eidaroos, K., Mehanny, S.
Influence of electrospinning parameters on biopolymers nanofibers, with emphasis on cellulose & chitosan
(2023) *Heliyon*, 9 (6).
- Zhong, T., Liu, W., Liu, H.
Green electrospinning of chitin propionate to manufacture nanofiber mats
(2021) *Carbohydr. Polym.*, 273.

- Avossa, J., Herwig, G., Toncelli, C., Itel, F., Rossi, R.M.
Electrospinning based on benign solvents: current definitions, implications and strategies
(2022) *Green Chem.*, 24 (6), pp. 2347-2375.
- Donato, R.K., Mija, A.
Keratin associations with synthetic, biosynthetic and natural polymers: an extensive review
(2019) *Polymers (Basel)*, 12 (1), p. 32.
- Al-Rajabi, M.M., Almanassra, I.W., Khalil, A.K.A., Atieh, M.A., Laoui, T., Khalil, K.A.
Facile coaxial electrospinning synthesis of polyacrylonitrile/cellulose acetate nanofiber membrane for oil-water separations
(2023) *Polymers (Basel)*, 15 (23).
- Zulkifli, M.Z.A., Nordin, D., Shaari, N., Kamarudin, S.K.
Overview of electrospinning for tissue engineering applications
(2023) *Polymers (Basel)*, 15 (11).
- Raza, S., Zhang, J., Ali, I., Li, X., Liu, C.
Recent trends in the development of biomass-based polymers from renewable resources and their environmental applications
(2020) *J. Taiwan Inst. Chem. Eng.*, 115, pp. 293-303.
- Huang, W.C., Zhao, D., Xue, C., Mao, X.
An efficient method for chitin production from crab shells by a natural deep eutectic solvent
(2022) *Mar. Life Sci. Technol.*, 4 (3), pp. 384-388.
- Ngasotter, S., Xavier, K.A.M., Meitei, M.M., Waikhom, D., Madhulika, Pathak, J., Singh, S.K.
Crustacean shell waste derived chitin and chitin nanomaterials for application in agriculture, food, and health – a review
(2023) *Carbohydr. Polym. Technol. Appl.*, 6.
- Wang, J., Teng, C., Yan, L.
Applications of deep eutectic solvents in the extraction, dissolution, and functional materials of chitin: research progress and prospects
(2022) *Green Chem.*, 24 (2), pp. 552-564.
- Jones, M., Kujundzic, M., John, S., Bismarck, A., Crab vs.
Mushroom: a review of crustacean and fungal chitin in wound treatment
(2020) *Mar. Drugs*, 18 (1).
- Zhong, Y., Cai, J., Zhang, L.-N.
A review of chitin solvents and their dissolution mechanisms
(2020) *Chin. J. Polym. Sci.*, 38 (10), pp. 1047-1060.
- Vicente, F.A., Ventura, S.P.M., Passos, H., Dias, A.C.R.V., Torres-Acosta, M.A., Novak, U., Likozar, B.
Crustacean waste biorefinery as a sustainable cost-effective business model
(2022) *Chem. Eng. J.*, 442.
- Hou, J., Aydemir, B.E., Dumanli, A.G.
Understanding the structural diversity of chitins as a versatile biomaterial
(2021) *Phil. Trans. R. Soc. A*, 379 (2206).
- Xiong, A., Ruan, L., Ye, K., Huang, Z., Yu, C.
Extraction of chitin from black soldier fly (*Hermetia illucens*) and its puparium by

- using biological treatment**
(2023) *Life (Basel)*, 13 (7).
- Kadokawa, J.
Application of ionic liquids for the functional materialization of chitin
(2022) *Mater. Adv.*, 3 (8), pp. 3355-3364.
 - Huang, J., Zhong, Y., Wei, P., Cai, J.
Rapid dissolution of β -chitin and hierarchical self-assembly of chitin chains in aqueous KOH/urea solution
(2021) *Green Chem.*, 23 (8), pp. 3048-3060.
 - Uto, T., Idenoue, S., Yamamoto, K., Kadokawa, J.I.
Understanding dissolution process of chitin crystal in ionic liquids: theoretical study
(2018) *Phys. Chem. Chem. Phys.*, 20 (31), pp. 20669-20677.
 - Knecht, E., Hibbert, E.
Some observations relating to chitin
(1926) *J. Soc. Dye. Colour.*, 42 (11), pp. 343-345.
 - Clark, G.L., Smith, A.F.
X-ray diffraction studies of chitin, chitosan and derivatives
(1936) *J. Phys. Chem.*, 40 (7), pp. 865-879.
 - Hackman, R.H.
Studies on chitin, V. The action of mineral acids on chitin
(1962) *Aust. J. Biol. Sci.*, 15 (3), pp. 526-537.
 - Yang, X., Liu, J., Pei, Y., Zheng, X., Tang, K.
Recent progress in preparation and application of nano-chitin materials
(2020) *Energy Environ. Mater.*, 3 (4), pp. 492-515.
 - Rokhati, N., Kusworo, T.D., Susanto, H., Widiassa, I.N., Aryanti, N., Adhiartha, A., Fahni, Y., Hamada, N.A.
Preparation of glucosamine by acid hydrolysis of chitin under microwave irradiation
(2020) *Proceedings of 2nd International Conference on Chemical Process and Product Engineering (ICCPPE) 2019*,
 - Wu, T., Wang, G., Gao, C., Chen, Z., Feng, L., Wang, P., Zeng, X., Wu, Z.
Phosphoric acid-based preparing of chitin nanofibers and nanospheres
(2015) *Cellulose*, 23 (1), pp. 477-491.
 - Tyshkunova, I.V., Chukhchin, D.G., Gofman, I.V., Pavlova, E.N., Ushakov, V.A., Vlasova, E.N., Poshina, D.N., Skorik, Y.A.
Chitin cryogels prepared by regeneration from phosphoric acid solutions
(2021) *Materials (Basel)*, 14 (18).
 - Ajavakom, A., Supsvetson, S., Somboot, A., Sukwattanasinitt, M.
Products from microwave and ultrasonic wave assisted acid hydrolysis of chitin
(2012) *Carbohydr. Polym.*, 90 (1), pp. 73-77.
 - Rojas, J., Madrigal, J., Ortiz, J.
Effect of acid hydrolysis on tableting properties of chitin obtained from shrimp heads
(2015) *Trop. J. Pharm. Res.*, 14 (7).
 - von Weimarn, P.P.
Conversion of fibroin,1 chitin, casein, and similar substances into the ropy-plastic state and colloidal solution2
(1927) *Ind. Eng. Chem.*, 19 (1), pp. 109-110.

- Gagnaire, D., Saint-Germain, J., Vincendon, M.
NMR studies of chitin and chitin derivatives
(1982) *Makromol. Chem.*, 183, pp. 593-601.
- Margoutidis, G., Johns, M.A., Kerton, F.M.
Dissolution studies of α -chitin fibers in freezing NaOH(aq)
(2021) *Cellulose*, 28 (4), pp. 1885-1891.
- Gorzędowski, J., Smiglak, M., Struszczyk, M.H., Puchalski, M., Krucińska, I.
Chitin fibre formation by the solution blow spinning method, using 1-butyl-3-methylimidazolium acetate ionic liquid as a solvent
(2020) *Fibres Text. East. Eur.*, 4 (142), pp. 42-48.
- Prasad, K., Murakami, M.A., Kaneko, Y., Takada, A., Nakamura, Y., Kadokawa, J.
Weak gel of chitin with ionic liquid, 1-allyl-3-methylimidazolium bromide
(2009) *Int. J. Biol. Macromol.*, 45 (3), pp. 221-225.
- Jaworska, M.M., Gorak, A.
Modification of chitin particles with chloride ionic liquids
(2016) *Mater. Lett.*, 164, pp. 341-343.
- Idenoue, S., Yamamoto, K., Kadokawa, J.
Dissolution of chitin in deep eutectic solvents composed of imidazolium ionic liquids and thiourea
(2019) *Chem. Eng.*, 3 (4).
- Ding, B., Cai, J., Huang, J., Zhang, L., Chen, Y., Shi, X., Du, Y., Kuga, S.
Facile preparation of robust and biocompatible chitin aerogels
(2012) *J. Mater. Chem.*, 22 (12).
- Liao, J., Huang, H.
A fungal chitin derived from *Herichium erinaceus* residue: dissolution, gelation and characterization
(2020) *Int. J. Biol. Macromol.*, 152, pp. 456-464.
- Ma, Q., Gao, X., Bi, X., Han, Q., Tu, L., Yang, Y., Shen, Y., Wang, M.
Dissolution and deacetylation of chitin in ionic liquid tetrabutylammonium hydroxide and its cascade reaction in enzyme treatment for chitin recycling
(2020) *Carbohydr. Polym.*, 230.
- Sharma, M., Mukesh, C., Mondal, D., Prasad, K.
Dissolution of α -chitin in deep eutectic solvents
(2013) *RSC Adv.*, 3 (39).
- Duan, B., Chang, C., Ding, B., Cai, J., Xu, M., Feng, S., Ren, J., Zhang, L.
High strength films with gas-barrier fabricated from chitin solution dissolved at low temperature
(2013) *J. Mater. Chem. A*, 1 (5), pp. 1867-1874.
- Huang, J., Zhong, Y., Lu, A., Zhang, L., Cai, J.
Temperature and time-dependent self-assembly and gelation behavior of chitin in aqueous KOH/urea solution
(2020) *Giant*, 4.
- Gong, P., Wang, J., Liu, B., Ru, G., Feng, J.
Dissolution of chitin in aqueous KOH
(2016) *Cellulose*, 23 (3), pp. 1705-1711.
- Zewude, D.A., Noguchi, T., Sato, K., Izawa, H., Ifuku, S.
Production of chitin nanoparticles by bottom-up approach from alkaline chitin

solution

(2022) *Int. J. Biol. Macromol.*, 210, pp. 123-127.

- Liu, Y., Liu, Z., Pan, W., Wu, Q.
Absorption behaviors and structure changes of chitin in alkali solution
(2008) *Carbohydr. Polym.*, 72 (2), pp. 235-239.
- Huang, J., Zhong, Y., Zhang, L., Cai, J.
Distinctive viewpoint on the rapid dissolution mechanism of α -chitin in aqueous potassium hydroxide-urea solution at low temperatures
(2020) *Macromolecules*, 53 (13), pp. 5588-5598.
- Li, F., You, X., Li, Q., Qin, D., Wang, M., Yuan, S., Chen, X., Bi, S.
Homogeneous deacetylation and degradation of chitin in NaOH/urea dissolution system
(2021) *Int. J. Biol. Macromol.*, 189, pp. 391-397.
- Chen, B., Sun, K., Zhang, K.
Rheological properties of chitin/lithium chloride, N,N-dimethyl acetamide solutions
(2004) *Carbohydr. Polym.*, 58 (1), pp. 65-69.
- de Vasconcelos, C.L., Bezerril, P.M., Pereira, M.R., Ginani, M.F., Fonseca, J.L.
Viscosity-temperature behavior of chitin solutions using lithium chloride/DMA as solvent
(2011) *Carbohydr. Res.*, 346 (5), pp. 614-618.
- Tokura, S., Nishimura, S.I., Sakairi, N., Nishi, N.
Biological activities of biodegradable polysaccharide
(1996) *Macromol. Symp.*, 101, pp. 389-396.
- Tamura, H., Nagahama, H., Tokura, S.
Preparation of chitin hydrogel under mild conditions
(2006) *Cellulose*, 13 (4), pp. 357-364.
- Feng, M., Lu, X., Hou, D., Zhang, S.
Solubility, Chain Characterization, and Derivatives of Chitin
(2020) *Preparation and Properties*, 1, pp. 101-129.
- Shamsuri, A.A., Abdan, K., Md Jamil, S.N.A.
Preparations and properties of ionic liquid-assisted electrospun biodegradable polymer fibers
(2022) *Polymers (Basel)*, 14 (12).
- Shamshina, J.L.
Chitin in ionic liquids: historical insights into the polymer's dissolution and isolation. A review
(2019) *Green Chem.*, 21 (15), pp. 3974-3993.
- Wu, Y., Sasaki, T., Irie, S., Sakurai, K.
A novel biomass-ionic liquid platform for the utilization of native chitin
(2008) *Polymer*, 49 (9), pp. 2321-2327.
- Walther, P., Ota, A., Müller, A., Hermanutz, F., Gähr, F., Buchmeiser, M.R.
Chitin foils and coatings prepared from ionic liquids
(2016) *Macromol. Mater. Eng.*, 301 (11), pp. 1337-1344.
- Kim, H., Kang, S., Li, K., Jung, D., Park, K., Lee, J.
Preparation and characterization of various chitin-glucan complexes derived from white button mushroom using a deep eutectic solvent-based ecofriendly method
(2021) *Int. J. Biol. Macromol.*, 169, pp. 122-129.

- Suthar, P., Kaushal, M., Vaidya, D., Thakur, M., Chauhan, P., Angmo, D., Kashyap, S., Negi, N.
Deep eutectic solvents (DES): an update on the applications in food sectors
(2023) *J. Agric. Food Res.*, 14.
- Wang, X., Zhou, P., Lv, X., Liang, Y.
Insight into the structure-function relationships of the solubility of chitin/chitosan in natural deep eutectic solvents
(2021) *Mater. Today Commun.*, 27.
- Cajnko, M.M., Vicente, F.A., Novak, U., Likozar, B.
Natural deep eutectic solvents (NaDES): translating cell biology to processing
(2023) *Green Chem.*, 25 (22), pp. 9045-9062.
- Ozel, N., Elibol, M.
A review on the potential uses of deep eutectic solvents in chitin and chitosan related processes
(2021) *Carbohydr. Polym.*, 262.
- Pacheco-Fernández, I., Pino, V.
Green solvents in analytical chemistry
(2019) *Curr. Opin. Green Sustain. Chem.*, 18, pp. 42-50.
- Wysokowski, M., Luu, R.K., Arevalo, S., Khare, E., Stachowiak, W., Niemczak, M., Jesionowski, T., Buehler, M.J.
Untapped potential of deep eutectic solvents for the synthesis of bioinspired inorganic-organic materials
(2023) *Chem. Mater.*, 35 (19), pp. 7878-7903.
- Mukesh, C., Mondal, D., Sharma, M., Prasad, K.
Choline chloride-thiourea, a deep eutectic solvent for the production of chitin nanofibers, carbohydrate
(2014) *Polymer*, 103, pp. 466-471.
- Vicente, F.A., Bradic, B., Novak, U., Likozar, B.
Alpha-chitin dissolution, N-deacetylation and valorization in deep eutectic solvents
(2020) *Biopolymers*, 111 (5).
- Min, B.M., Lee, S.W., Lim, J.N., You, Y., Lee, T.S., Kang, P.H., Park, W.H.
Chitin and chitosan nanofibers: electrospinning of chitin and deacetylation of chitin nanofibers
(2004) *Polymer*, 45 (21), pp. 7137-7142.
- Zhong, C., Cooper, A., Kapetanovic, A., Fang, Z., Zhang, M., Rolandi, M.
A facile bottom-up route to self-assembled biogenic chitin nanofibers
(2010) *Soft Matter*, 6 (21), pp. 5298-5301.
- Zhou, N., Yang, P., Chen, J., Wei, G., Wang, C., Zhang, A., Chen, K., Ouyang, P.
Effect of organic solvents treatment on structure of chitin and its enzymatic hydrolysis
(2022) *Polym. Degrad. Stab.*, 198.
- Arshad, M., Zubair, M., Ullah, A.
Miscibility, Properties, and Biodegradability of Chitin and Chitosan
(2020), pp. 377-399.
- Mejía Agüero, L.E., Saul, C.K., de Freitas, R.A., Rabello Duarte, M.E., Nosedá, M.D.
Electrospinning of marine polysaccharides: processing and chemical aspects, challenges, and future prospects
(2022) *Nanotechnol. Rev.*, 11 (1), pp. 3250-3280.

- Antaby, E., Klinkhammer, K., Sabantina, L.
Electrospinning of chitosan for antibacterial applications—current trends
(2021) *Appl. Sci.*, 11 (24), p. 11937.
- Das, K.P., Sharma, D., Satapathy, B.K.
Electrospun fibrous constructs towards clean and sustainable agricultural prospects: SWOT analysis and TOWS based strategy assessment
(2022) *J. Clean. Prod.*, 368.
- Munteanu, B.S., Vasile, C.
Encapsulation of natural bioactive compounds by electrospinning-applications in food storage and safety
(2021) *Polymers (Basel)*, 13 (21).
- Li, H., Chen, X., Lu, W., Wang, J., Xu, Y., Guo, Y.
Application of electrospinning in antibacterial field
(2021) *Nanomaterials (Basel)*, 11 (7), p. 1822.
- Rathore, P., Schiffman, J.D.
Beyond the single-nozzle: coaxial electrospinning enables innovative nanofiber chemistries, geometries, and applications
(2021) *ACS Appl. Mater. Interfaces*, 13 (1), pp. 48-66.
- Han, T., Yarin, A.L., Reneker, D.H.
Viscoelastic electrospun jets: initial stresses and elongational rheometry
(2008) *Polymer*, 49 (6), pp. 1651-1658.
- Haider, A., Haider, S., Kang, I.-K.
A comprehensive review summarizing the effect of electrospinning parameters and potential applications of nanofibers in biomedical and biotechnology
(2018) *Arab. J. Chem.*, 11 (8), pp. 1165-1188.
- Haider, S., Al-Zeghayer, Y., Ahmed Ali, F.A., Haider, A., Mahmood, A., Al-Masry, W.A., Imran, M., Aijaz, M.O.
Highly aligned narrow diameter chitosan electrospun nanofibers
(2013) *J. Polym. Res.*, 20 (105).
- Kim, S.C., Kang, S., Lee, H., Kwak, D.-B., Ou, Q., Pei, C., Pui, D.Y.H.
Nanofiber filter performance improvement: nanofiber layer uniformity and branched nanofiber
(2020) *Aerosol Air Qual. Res.*, 20 (1), pp. 80-88.
- Huan, S., Liu, G., Han, G., Cheng, W., Fu, Z., Wu, Q., Wang, Q.
Effect of experimental parameters on morphological, mechanical and hydrophobic properties of electrospun polystyrene fibers
(2015) *Materials*, 8 (5), pp. 2718-2734.
- Pelipenko, J., Kristl, J., Jankovic, B., Baumgartner, S., Kocbek, P.
The impact of relative humidity during electrospinning on the morphology and mechanical properties of nanofibers
(2013) *Int. J. Pharm.*, 456 (1), pp. 125-134.
- Metreveli, G., Wågberg, L., Emmoth, E., Belák, S., Strømme, M., Mihranyan, A.
A size-exclusion nanocellulose filter paper for virus removal
(2014) *Adv. Healthc. Mater.*, 3, pp. 1546-1550.
- Chen, S., John, J.V., McCarthy, A., Xie, J.
New forms of electrospun nanofiber materials for biomedical applications
(2020) *J. Mater. Chem. B*, 8 (17), pp. 3733-3746.

- Park, K.E., Jung, S.Y., Lee, S.J., Min, B.M., Park, W.H.
Biomimetic nanofibrous scaffolds: preparation and characterization of chitin/silk fibroin blend nanofibers
(2006) *Int. J. Biol. Macromol.*, 38 (3-5), pp. 165-173.
- Saudi, S., Jun, S., Fialkova, S., Surendran, V., Chandrasekaran, A., Bhattarai, S.R., Sankar, J., Bhattarai, N.
Incorporating nanoconfined chitin-fibrils in poly (epsilon-caprolactone) membrane scaffolds improves mechanical and chemical properties for biomedical application
(2023) *J. Biomed. Mater. Res. A*, 111 (8), pp. 1185-1199.
- Jung, H.S., Kim, M.H., Shin, J.Y., Park, S.R., Jung, J.-Y., Park, W.H.
Electrospinning and wound healing activity of β -chitin extracted from cuttlefish bone
(2018) *Carbohydr. Polym.*, 193, pp. 205-211.
- Shirazi, M.M.A., Bazgir, S., Meshkani, F.
Electrospun nanofibrous membranes for water treatment
(2020) *Adv. Membr. Technol.*, 57 (3), pp. 467-504.
- Marinho, B.A., de Souza, S.M.A.G.U., de Souza, A.A.U., Hotza, D.
Electrospun TiO₂ nanofibers for water and wastewater treatment: a review
(2020) *J. Mater. Sci.*, 56 (9), pp. 5428-5448.
- Cui, J., Li, F., Wang, Y., Zhang, Q., Ma, W., Huang, C.
Electrospun nanofiber membranes for wastewater treatment applications
(2020) *Sep. Purif. Technol.*, 250.
- Nayl, A.A., Abd-Elhamid, A.I., Awwad, N.S., Abdelgawad, M.A., Wu, J., Mo, X., Gomha, S.M., Brase, S.
Review of the recent advances in electrospun nanofibers applications in water purification
(2022) *Polymers (Basel)*, 14 (8).
- Tang, Y., Cai, Z., Sun, X., Chong, C., Yan, X., Li, M., Xu, J.
Electrospun nanofiber-based membranes for water treatment
(2022) *Polymers (Basel)*, 14 (10).
- Gopi, S., Balakrishnan, P., Pius, A., Thomas, S.
Chitin nanowhisker (ChNW)-functionalized electrospun PVDF membrane for enhanced removal of indigo carmine
(2017) *Carbohydr. Polym.*, 165, pp. 115-122.
- Li, Q.H., Dong, M., Li, R., Cui, Y.Q., Xie, G.X., Wang, X.X., Long, Y.Z.
Enhancement of Cr(VI) removal efficiency via adsorption/photocatalysis synergy using electrospun chitosan/g-C₃N₄/TiO₂ nanofibers
(2021) *Carbohydr. Polym.*, 253.
- Talukder, M.E., Pervez, M.N., Jianming, W., Gao, Z., Stylios, G.K., Hassan, M.M., Song, H., Naddeo, V.
Chitosan-functionalized sodium alginate-based electrospun nanofiber membrane for As (III) removal from aqueous solution
(2021) *J. Environ. Chem. Eng.*, 9 (6).
- Song, R., Yan, J., Xu, S., Wang, Y., Ye, T., Chang, J., Deng, H., Li, B.
Silver ions/ovalbumin films layer-by-layer self-assembled polyacrylonitrile nanofibrous mats and their antibacterial activity
(2013) *Colloids Surf. B: Biointerfaces*, 108, pp. 322-328.
- Zhang, M., Ahmed, A., Xu, L.
Electrospun nanofibers for functional food packaging application

- (2023) *Materials (Basel)*, 16 (17).
- Neo, Y.P., Ray, S., Perera, C.O.
Fabrication of Functional Electrospun Nanostructures for Food Applications
(2018) *Role of Materials Science in Food Bioengineering*, pp. 109-146.
 - Duan, M., Yu, S., Sun, J., Jiang, H., Zhao, J., Tong, C., Hu, Y., Wu, C.
Development and characterization of electrospun nanofibers based on pullulan/chitin nanofibers containing curcumin and anthocyanins for active-intelligent food packaging
(2021) *Int. J. Biol. Macromol.*, 187, pp. 332-340.
 - Duan, M., Sun, J., Huang, Y., Jiang, H., Hu, Y., Pang, J., Wu, C.
Electrospun gelatin/chitosan nanofibers containing curcumin for multifunctional food packaging
(2023) *Food Sci. Human Wellness*, 12 (2), pp. 614-621.
 - Li, S., Yan, Y., Guan, X., Huang, K.
Preparation of a hordein-quercetin-chitosan antioxidant electrospun nanofibre film for food packaging and improvement of the film hydrophobic properties by heat treatment
(2020) *Food Packag. Shelf Life*, 23.
 - Qin, X.
Coaxial electrospinning of nanofibers
(2017) *Woodhead Publishing Series in Textiles*, pp. 41-71.
Woodhead Publishing
 - Hu, Q.-R., Shen, X.-Y., Qian, X.-M., Huang, G.-Y., Yuan, M.-Q.
The personal protective equipment (PPE) based on individual combat: a systematic review and trend analysis
(2023) *Def. Technol.*, 28, pp. 195-221.
 - Khan, M.K.R.
CBRN personal protective clothing
(2021),
(accessed 26 February 2024)
 - Baji, A., Agarwal, K., Oopath, S.V.
Emerging developments in the use of electrospun fibers and membranes for protective clothing applications
(2020) *Polymers (Basel)*, 12 (2).
 - Ding, B., Yu, J.
Electrospun Nanofibers for Energy and Environmental Applications. Nanostructure Science and Technology
(2014), Springer Berlin Heidelberg
 - Agarwal, S., Greiner, A., Wendorff, J.H.
Functional materials by electrospinning of polymers
(2013) *Prog. Polym. Sci.*, 38 (6), pp. 963-991.
 - Yezer, I., Demirkol, D.O.
Cellulose acetate-chitosan based electrospun nanofibers for bio-functionalized surface design in biosensing
(2020) *Cellulose*, 27 (17), pp. 10183-10197.
 - Enculescu, M., Costas, A., Evanghelidis, A., Enculescu, I.
Fabrication of ZnO and TiO(2) nanotubes via flexible electro-spun nanofibers for photocatalytic applications
(2021) *Nanomaterials (Basel)*, 11 (5).

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