





Whole-Genome Sequence of a *Stenotrophomonas maltophilia* Isolate from Tap Water in an Intensive Care Unit

Ummu Afeera Zainulabid, a,b Shing Wei Siew, Siti Munirah Musa, Sharmeen Nellisa Soffian, Petrick Periyasamy, Bhajar Fauzan Ahmad Shangar Fauzan Ahmad Shanga

^aDepartment of Internal Medicine, Kulliyyah of Medicine, International Islamic University Malaysia, Kuantan, Pahang, Malaysia

ABSTRACT Here, we present a 4,508,936-bp complete genome sequence of *Stenotro-phomonas maltophilia* strain HW002Y, which was isolated from the tap water in an intensive care unit at Sultan Ahmad Shah Medical Centre at the International Islamic University of Malaysia (Kuantan, Pahang, Malaysia). Sequencing was performed using a Nanopore Flongle flow cell.

he Gram-negative bacillus Stenotrophomonas maltophilia is a multidrug-resistant organism that commonly infects patients in intensive care units. The source of contact is from environmental water sources, such as hospital tap water, direct contact, ingestion, aspiration, or aerosolization (1, 2). A sample was isolated from the hospital tap water in an intensive care unit at Sultan Ahmad Shah Medical Centre at the International Islamic University of Malaysia (IIUM) (Kuantan, Pahang, Malaysia). Three liters of tap water was vacuum filtered to concentrate the bacteria. The membrane filter was transferred to a collection tube containing phosphate-buffered saline, vortex-mixed, and left to soak overnight at room temperature. Subsequently, 0.1 mL of the solution was taken and cultured overnight at 37°C on ampicillin-containing Luria-Bertani agar. A presumed colony of S. maltophilia was selected, and the genomic DNA (gDNA) of the strain was extracted as described previously, using the GenElute bacterial gDNA kit (Sigma-Aldrich Co., St. Louis, MO, USA) (3). A high-sensitivity kit (DeNovix, Wilmington, DE) and agarose gel electrophoresis were used to evaluate the concentration and quality of the qDNA, respectively. Prior to library preparation, DNA was fragmented to an average length of >6 kb using SPRIselect reagent (Beckman Coulter Inc., Indianapolis, IN, USA) as described previously (4). Briefly, 1.25 µg of DNA was used for library preparation with a ligation sequencing kit (SQK-LSK110; Oxford Nanopore Technologies, Oxford, UK). The prepared library was then loaded onto a Flongle flow cell (R9.4.1; Oxford Nanopore Technologies) for 72 h. Base calling of raw data was performed using Guppy v6.0.6 (super accurate model). The raw read length of the genome is 12,000 reads and the N_{50} value is 14,704 bp. The base-called reads were length filtered to include only reads with a minimum read length of 3 kb, followed by de novo assembly using Flye v2.9 (nano-hq model) (5). The assembled genome was subsequently polished with one round with Racon v1.4.3 (https://github.com/ isovic/racon) and then one round with medaka v1.6.0 (https://github.com/nanoporetech/ medaka) (6), generating the final consensus assembly, which consists of one circular contig, as assessed by Flye v2.9 (7). HW002Y has a total length of 4,508,936 bp, with an average GC content of 66.6% and genome coverage of 231.0×. The percent identity between the query sequences and the bacterial species discovered with autoMLST (8) was examined via the Average Nucleotide Identity (ANI) Calculator v1.0 (https://www.ezbiocloud.net/tools/ani) (9), which showed that the sequence was 97.97% identical to the S. maltophilia reference genome (GenBank assembly accession number GCF_001274655). Due to the high level of nucleotide identity, it is therefore proven that the isolated bacterial sample is S. maltophilia.

Editor Catherine Putonti, Loyola University Chicago

Copyright © 2023 Zainulabid et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Ummu Afeera Zainulabid, ummuafeera@iium.edu.my, or Hajar Fauzan Ahmad,

fauzanahmad@ump.edu.my.

The authors declare no conflict of interest.

Received 18 September 2022 **Accepted** 22 December 2022 **Published** 12 January 2023

^bMedical Department, Faculty of Medicine, Universiti Kebangsaan Malaysia, Cheras, Kuala Lumpur, Malaysia

^cFaculty of Industrial Sciences and Technology, Universiti Malaysia Pahang, Gambang, Pahang, Malaysia

Downloaded from https://journals.asm.org/journal/mra on 27 May 2025 by 210.48.218.3.

Genome annotation was carried out using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) v6.2 with the GeneMarkS2 v1.14_1.25 method (10), which resulted in one contig with 4,151 coding sequences and 85 RNAs. Unless otherwise noted, default parameters were used for all software tools.

The study was approved by the IIUM Kulliyyah of Medicine Research Committee (KRC) (research identification number 801) and the Department of Education and Research, Sultan Ahmad Shah Medical Centre, IIUM (registration number IISR22-06).

Data availability. The complete genome sequencing reads for *Stenotrophomonas maltophilia* HW002Y are available in the NCBI SRA under accession number SRR19579117 and BioProject accession number PRJNA847001. The GenBank accession number is CP104169.1, and the assembly accession number is GCA 025200825.1.

ACKNOWLEDGMENTS

This research was supported in full by a Kurita Asia Research Grant (grant 21Pmy012-22I) provided by the Kurita Water and Environment Foundation. The funders had no role in study design, data collection and analysis, publishing decisions, or manuscript preparation.

REFERENCES

- Guerci P, Bellut H, Mokhtari M, Gaudefroy J, Mongardon N, Charpentier C, Louis G, Tashk P, Dubost C, Ledochowski S, Kimmoun A, Godet T, Pottecher J, Lalot JM, Novy E, Hajage D, Bouglé A, Constantin JM, Perbet S, AZUREA Research Network. 2019. Outcomes of Stenotrophomonas maltophilia hospitalacquired pneumonia in intensive care unit: a nationwide retrospective study. Crit Care 23:371. https://doi.org/10.1186/s13054-019-2649-5.
- Cervia JS, Ortolano GA, Canonica FP. 2008. Hospital tap water as a source of *Stenotrophomonas maltophilia* infection. Clin Infect Dis 46:1485–1487. https://doi.org/10.1086/587180.
- Ahmad HF, Schreiber L, Marshall IPG, Andersen PJ, Castro-Mejía JL, Nielsen DS. 2019. Draft genome sequence of *Streptococcus anginosus* strain CALM001, isolated from the gut of an elderly Dane. Microbiol Resour Announc 8:e00379-19. https://doi.org/10.1128/MRA.00379-19.
- Zainulabid UA, Mohamad Zain N, Arumugam J, Kamarudin N, Zainal Abidin MA, Abd Mokti AS, Nordin N, Faizal Rakawi A, Abdul Majid AS, Ashok G, Francis AL, Tay DD, Vijayalakshami N, Hin HS, Ahmad HF. 2022. Near-complete whole-genome sequencing of two *Burkholderia pseudomallei* strains harbouring novel molecular class D beta-lactamase genes, isolated from Malaysia. Microbiol Resour Announc 11:e00468-22. https://doi.org/10.1128/mra.00468-22.

- Kolmogorov M, Yuan J, Lin Y, Pevzner PA. 2019. Assembly of long, error-prone reads using repeat graphs. Nat Biotechnol 37:540–546. https://doi.org/10.1038/ s41587-019-0072-8.
- Vaser R, Sović I, Nagarajan N, Šikić M. 2017. Fast and accurate de novo genome assembly from long uncorrected reads. Genome Res 27:737–746. https://doi.org/10.1101/gr.214270.116.
- Kolmogorov M, Bickhart DM, Behsaz B, Gurevich A, Rayko M, Shin SB, Kuhn K, Yuan J, Polevikov E, Smith TPL, Pevzner PA. 2020. metaFlye: scalable long-read metagenome assembly using repeat graphs. Nat Methods 17:1103–1110. https://doi.org/10.1038/s41592-020-00971-x.
- Alanjary M, Steinke K, Ziemert N. 2019. AutoMLST: an automated web server for generating multi-locus species trees highlighting natural product potential. Nucleic Acids Res 47:W276–W282. https://doi.org/10.1093/nar/qkz282.
- Yoon SH, Ha S, Lim J, Kwon S, Chun J. 2017. A large-scale evaluation of algorithms to calculate average nucleotide identity. Antonie Van Leeuwenhoek 110:1281–1286. https://doi.org/10.1007/s10482-017-0844-4.
- Lomsadze A, Gemayel K, Tang S, Borodovsky M. 2018. Modeling leaderless transcription and atypical genes results in more accurate gene prediction in prokaryotes. Genome Res 28:1079–1089. https://doi.org/10.1101/gr.230615.117.